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(54) Title: OGRI GENE DISRUPTIONS, COMPOSITIONS AND METHODS RELATING THERETO

(57) Abstract: The present invention relates to transgenic animals, as well as compositions and methods relating to the characterization of gene function. Specifically, the present invention provides transgenic mice comprising mutations in an OGRI gene. Such transgenic mice are useful as models for disease and for identifying agents that modulate gene expression and gene function, and as potential treatments for various disease states and disease conditions.

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## OGR1 GENE DISRUPTIONS, COMPOSITIONS AND METHODS RELATING THERETO

### Field of the Invention

The present invention relates compositions, including transgenic animals and methods  
5 relating to the characterization of gene function.

### Background of the Invention

G-protein-coupled receptors (GPCRs) are an important family of cell-surface receptors. Many of these receptors have been identified by homology cloning or by expression cloning using ligand-binding or cell-activation properties to identify them. GPCRs mediate cellular responses to  
10 diverse signaling molecules, including hormones, neurotransmitters, and local mediators. These signaling molecules vary in their structure and function, and include proteins, small peptides, amino acid and fatty acid derivatives. The GPCRs, however, have similar structure, a transmembrane seven-helix protein (7TM) domain and are almost certainly evolutionarily related. (For a review, *see e.g.*, Alberts *et al.*, *Molecular Biology of the Cell*, 3<sup>rd</sup> edition, p. 734-759).

15 There is an enormous therapeutic interest in manipulating or modulating (either enhancing or suppressing) GPCR signal transduction. GPCRs constitute the most prominent family of validated drug targets within biomedical research. Much progress has been made in understanding the mechanisms of action of these key proteins and their physiological functions. The *in vivo* manipulation of GPCRs using transgenic and gene knockout approaches have been particularly  
20 successful in assessing the roles of GPCRs in animal and human physiology. Drug discovery efforts are focused on producing highly specific compounds based on subtle definition of receptor subtypes, and new therapeutic opportunities may be provided by investigation of orphan receptors whose natural ligands are unidentified.

A novel GPCR named ovarian cancer GPCR (OGR1; aka GPR68) was cloned from an  
25 ovarian cancer cell line (GenBank Accession No.: U48405; GI No.: 1457938; Xu and Casey, *Genomics* 35(2):397-402 (1996)). The predicted open reading frame of OGR1 encodes a protein of 365 amino acids and shares strongest homology with the orphan receptor GPR4. OGR1 is believed to be expressed as a single 3.0-kb transcript in several tissues, including spleen, testis, small intestine, peripheral blood leukocytes, brain, heart, lung, placenta, and kidney, with no detectable OGR1  
30 expression in thymus, prostate, ovary, colon, liver, skeletal muscle, or pancreas.

Given the importance of GPCRs in biological and disease processes, a clear need exists for *in vivo* characterization of GPCRs, in particular, OGR1, which may aid in the identification and discovery of therapeutics and treatments useful in preventing, ameliorating or correcting dysfunctions or diseases.

### Summary of the Invention

The present invention generally relates to transgenic animals, as well as to compositions and methods relating to the characterization of gene function.

The present invention provides transgenic cells comprising a disruption in an OGR1 gene.

5 The transgenic cells of the present invention are comprised of any cells capable of undergoing homologous recombination. Preferably, the cells of the present invention are stem cells and more preferably, embryonic stem (ES) cells, and most preferably, murine ES cells. According to one embodiment, the transgenic cells are produced by introducing a targeting construct into a stem cell to produce a homologous recombinant, resulting in a mutation of the OGR1 gene. In another  
10 embodiment, the transgenic cells are derived from the transgenic animals described below. The cells derived from the transgenic animals includes cells that are isolated or present in a tissue or organ, and any cell lines or any progeny thereof.

The present invention also provides a targeting construct and methods of producing the targeting construct that when introduced into stem cells produces a homologous recombinant. In one  
15 embodiment, the targeting construct of the present invention comprises first and second polynucleotide sequences that are homologous to the OGR1 gene. The targeting construct may also comprise a polynucleotide sequence that encodes a selectable marker that is preferably positioned between the two different homologous polynucleotide sequences in the construct. The targeting construct may also comprise other regulatory elements that can enhance homologous recombination.

20 The present invention further provides non-human transgenic animals and methods of producing such non-human transgenic animals comprising a disruption in an OGR1 gene. The transgenic animals of the present invention include transgenic animals that are heterozygous and homozygous for a null mutation in the OGR1 gene. In one aspect, the transgenic animals of the present invention are defective in the function of the OGR1 gene.

25 In another aspect, the transgenic animals of the present invention comprise a phenotype associated with having a mutation in an OGR1 gene. Preferably, the transgenic animals are rodents and, most preferably, are mice.

In a preferred embodiment, the present invention provides a transgenic mouse comprising a disruption in an OGR1 gene, wherein there is no native expression of the endogenous OGR1 gene.

30 In another preferred embodiment, the transgenic mice of the present invention exhibit hypoactivity, as characterized by a decrease in the total distance traveled in an open field test. The hypoactivity is the opposite behavior as that seen in human hyperactivity disorders, such as attention deficit hyperactivity disorder (ADHD), attention deficit disorder (ADD), hyperactive child syndrome, minimal brain dysfunction, or hyperactivity. As such, the present invention provides methods, cells  
35 and transgenic mice useful in the discovery of treatments for such disorders.

In yet another embodiment, the transgenic mice of the present invention exhibit a motor abnormality. In accordance with this embodiment, the motor abnormality may comprise impaired motor coordination, impaired balance, or ataxia. In a preferred embodiment, the motor abnormality is characterized by a decrease in the speed at which the transgenic mice fall from a rotating rod in a rotarod test.

In yet another embodiment, the transgenic mice of the present invention exhibit a stimulus processing abnormality. In one aspect, the stimulus processing abnormality is a stimulus processing deficit, as characterized by a decrease in the response to startle stimuli in a startle response test. In another aspect, the stimulus processing abnormality is enhanced prepulse inhibition, as characterized by an increase in startle gating seen in startle response testing. Accordingly, the prepulse inhibition as seen in the transgenic mice is the opposite of a processing deficit seen in schizophrenic patients.

In one aspect of the present invention, a transgenic mouse having a disruption in the OGR1 gene exhibits a phenotype consistent with one or more symptoms of a disease associated with OGR1. In a preferred embodiment, a transgenic mouse having a disruption in the OGR1 gene exhibits a phenotype consistent with human hyperactivity. In another preferred embodiment, a transgenic mouse having a disruption in the OGR1 gene exhibits a phenotype consistent with human schizophrenia.

The transgenic mice of the present invention may be used as an *in vivo* model to study various disease states or conditions in which OGR1 may be implicated or may be involved, such as motor coordination, balance, ataxia, hyperactivity, or schizophrenia. The transgenic mice of the present invention may also be used to evaluate various treatments or to identify agents for the treatment of disease states or conditions in which OGR1 may be implicated or may be involved, such as motor coordination, balance, ataxia, hyperactivity, or schizophrenia. In addition, cells comprising a disruption in the OGR1 gene, including cells derived from the transgenic animals of the present invention, may also be used in the study of or to evaluate or identify treatments for disease states or conditions in which OGR1 may be implicated, such as motor coordination, balance, ataxia, hyperactivity, or schizophrenia.

The present invention also provides methods of identifying agents capable of affecting a phenotype of a transgenic animal. For example, a putative agent is administered to the transgenic animal and a response of the transgenic animal to the putative agent is measured and compared to the response of a "normal" or wild-type mouse, or alternatively compared to a transgenic animal control (without agent administration). The invention further provides agents identified according to such methods. The present invention also provides methods of identifying agents useful as therapeutic agents for treating conditions associated with a disruption or other mutation (including naturally occurring mutations) of the OGR1 gene.

One aspect of the present invention relates to a method of identifying a potential therapeutic agent for the treatment of a disease associated with the OGR1 gene, in which the method includes the steps of administering the potential therapeutic agent to a transgenic mouse having a disruption in an OGR1 gene and determining whether the potential therapeutic agent modulates the disease associated with the OGR1 gene, wherein the modulation of the disease identifies a potential therapeutic agent for the treatment of that disease. In a preferred embodiment, the disease associated with the OGR1 gene is hyperactivity. In another preferred embodiment, the disease associated with the OGR1 gene is schizophrenia. Accordingly, the foregoing method may be used to identify potential therapeutic agents for the treatment of hyperactivity or schizophrenia.

A further aspect of the present invention provides a method of identifying a potential therapeutic agent for the treatment of a disease associated with the OGR1 gene, in which the method includes the steps of contacting the potential therapeutic agent with OGR1 gene product and determining whether the potential therapeutic agent modulates that product, wherein modulation of the gene product identifies a potential therapeutic agent for the treatment of the disease associated with the OGR1 gene. In a preferred embodiment, the disease associated with the OGR1 gene is hyperactivity. In another preferred embodiment, the disease associated with the OGR1 gene is schizophrenia. In accordance with these embodiments, the foregoing method may be used to identify potential therapeutic agents for the treatment of hyperactivity or schizophrenia.

The present invention further provides a method of identifying agents having an effect on OGR1 expression or function. The method includes administering an effective amount of the agent to a transgenic animal, preferably a mouse. The method includes measuring a response of the transgenic animal, for example, to the agent, and comparing the response of the transgenic animal to a control animal, which may be, for example, a wild-type animal or alternatively, a transgenic animal control. Compounds that may have an effect on OGR1 expression or function may also be screened against cells in cell-based assays, for example, to identify such compounds.

The invention also provides cell lines comprising nucleic acid sequences of an OGR1 gene. Such cell lines may be capable of expressing such sequences by virtue of operable linkage to a promoter functional in the cell line. Preferably, expression of the OGR1 gene sequence is under the control of an inducible promoter. Also provided are methods of identifying agents that interact with the OGR1 gene, comprising the steps of contacting the OGR1 gene with an agent and detecting an agent/OGR1 gene complex. Such complexes can be detected by, for example, measuring expression of an operably linked detectable marker.

The invention further provides methods of treating diseases or conditions associated with a disruption in an OGR1 gene, and more particularly, to a disruption or other alteration in the expression or function of the OGR1 gene. In a preferred embodiment, methods of the present invention involve treating diseases or conditions associated with a disruption or other alteration in the

OGR1 gene's expression or function, including administering to a subject in need, a therapeutic agent that affects OGR1 expression or function. In accordance with this embodiment, the method comprises administration of a therapeutically effective amount of a natural, synthetic, semi-synthetic, or recombinant OGR1 gene, OGR1 gene products or fragments thereof as well as natural, synthetic, 5 semi-synthetic or recombinant analogs.

In one aspect of the present invention, a therapeutic agent for treating a disease associated with the OGR1 gene modulates the OGR1 gene product. Another aspect of the present invention relates to a therapeutic agent for treating a disease associated with the OGR1 gene, in which the agent is an agonist or antagonist of the OGR1 gene product. In a further aspect of the present invention, a 10 therapeutic agent for treating hyperactivity is provided that modulates the OGR1. In a preferred embodiment, the therapeutic agent for treating hyperactivity is an antagonist of OGR1. The present invention further relates to a therapeutic agent for treating schizophrenia, wherein the agent modulates OGR1. In a preferred embodiment, the therapeutic agent for treating schizophrenia is an antagonist of OGR1.

15 The present invention also provides compositions comprising or derived from ligands or other molecules or compounds that bind to or interact with OGR1, including agonists or antagonists of OGR1. Such agonists or antagonists of OGR1 include antibodies and antibody mimetics, as well as other molecules that can readily be identified by routine assays and experiments well known in the art.

The present invention further provides methods of treating diseases or conditions associated 20 with disrupted targeted gene expression or function, wherein the methods comprise detecting and replacing through gene therapy mutated or otherwise defective or abnormal OGR1 genes.

### Definitions

The following terms have the meanings ascribed to them below unless specified otherwise.

25 The term "gene" refers to (a) a gene containing at least one of the DNA sequences disclosed herein; (b) any DNA sequence that encodes the amino acid sequence encoded by the DNA sequences disclosed herein and/or; (c) any DNA sequence that hybridizes to the complement of the coding sequences disclosed herein. Preferably, the term includes coding as well as noncoding regions, and preferably includes all sequences necessary for normal gene expression including promoters, enhancers and other regulatory sequences.

30 The terms "polynucleotide" and "nucleic acid molecule" are used interchangeably to refer to polymeric forms of nucleotides of any length. The polynucleotides may contain deoxyribonucleotides, ribonucleotides and/or their analogs. Nucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The term "polynucleotide" includes single-, double-stranded and triple helical molecules.

35 "Oligonucleotide" refers to polynucleotides of between 5 and about 100 nucleotides of single- or double-stranded DNA. Oligonucleotides are also known as oligomers or oligos and may be

isolated from genes, or chemically synthesized by methods known in the art. A "primer" refers to an oligonucleotide, usually single-stranded, that provides a 3'-hydroxyl end for the initiation of enzyme-mediated nucleic acid synthesis. The following are non-limiting embodiments of polynucleotides: a gene or gene fragment, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant  
5 polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes and primers. A nucleic acid molecule may also comprise modified nucleic acid molecules, such as methylated nucleic acid molecules and nucleic acid molecule analogs. Analogs of purines and pyrimidines are known in the art, and include, but are not limited to, aziridinycytosine, 4-acetylcytosine, 5-fluorouracil, 5-bromouracil, 5-carboxymethylamino-  
10 methyl-2-thiouracil, 5-carboxymethyl-aminomethyluracil, inosine, N6-isopentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, pseudouracil, 5-pentyluracil and 2,6-diaminopurine. The use of uracil as a substitute for thymine in a deoxyribonucleic acid is also considered an analogous form of pyrimidine.

15 A "fragment" of a polynucleotide is a polynucleotide comprised of at least 9 contiguous nucleotides, preferably at least 15 contiguous nucleotides and more preferably at least 45 nucleotides, of coding or non-coding sequences.

The term "gene targeting" refers to a type of homologous recombination that occurs when a fragment of genomic DNA is introduced into a mammalian cell and that fragment locates and  
20 recombines with endogenous homologous sequences.

The term "homologous recombination" refers to the exchange of DNA fragments between two DNA molecules or chromatids at the site of homologous nucleotide sequences.

The term "homologous" as used herein denotes a characteristic of a DNA sequence having at least about 70 percent sequence identity as compared to a reference sequence, typically at least about  
25 85 percent sequence identity, preferably at least about 95 percent sequence identity, and more preferably about 98 percent sequence identity, and most preferably about 100 percent sequence identity as compared to a reference sequence. Homology can be determined using, for example, a "BLASTN" algorithm. It is understood that homologous sequences can accommodate insertions, deletions and substitutions in the nucleotide sequence. Thus, linear sequences of nucleotides can be  
30 essentially identical even if some of the nucleotide residues do not precisely correspond or align. The reference sequence may be a subset of a larger sequence, such as a portion of a gene or flanking sequence, or a repetitive portion of a chromosome.

The term "target gene" (alternatively referred to as "target gene sequence" or "target DNA sequence" or "target sequence") refers to any nucleic acid molecule, polynucleotide, or gene to be  
35 modified by homologous recombination. The target sequence includes an intact gene, an exon or intron, a regulatory sequence or any region between genes. The target gene may comprise a portion

of a particular gene or genetic locus in the individual's genomic DNA. As provided herein, the target gene of the present invention is an OGR1 gene, or a homolog or ortholog thereof.

"OGR1" comprises any one of the following: (1) the sequence shown in Figure 1 (SEQ ID NO:1) or identified in GenBank as Accession No.: U48405; GI No.: 1457938; (2) the OGR1 protein  
5 as shown in Figure 2 (SEQ ID NO:2) or identified in GenBank Accession No.: AAC50596; GI No.: 1457939; or (3) any homologues of the above identified sequences.

The term "OGR1 molecule" refers to OGR1 as defined above or variants, derivatives, active fragments or mutants of OGR1.

As used herein, a "variant" of OGR1 is defined as an amino acid sequence that is different by  
10 one or more amino acid substitutions. The variant may have "conservative" changes, wherein a substituted amino acid has similar structural or chemical properties, *e.g.*, replacement of a leucine with isoleucine. More rarely, a variant may have "nonconservative" changes, *e.g.*, replacement of a glycine with a tryptophan. Similar minor variations may also include amino acid deletions or insertions, or both. Guidance in determining which and how many amino acid residues may be  
15 substituted, inserted or deleted without abolishing biological or immunological activity may be found using computer programs well known in the art, for example, DNASTar software.

The term "active fragment" refers to a fragment of an OGR1 that is biologically or immunologically active. The term "biologically active" refers to an OGR1 having structural, regulatory or biochemical functions of the naturally occurring OGR1. Likewise, "immunologically  
20 active" defines the capability of the natural, recombinant or synthetic OGR1, or any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

The term "derivative", as used herein, refers to the chemical modification of a nucleic acid sequence encoding an OGR1 or the encoded OGR1 protein. An example of such modifications would  
25 be replacement of hydrogen by an alkyl, acyl, or amino group. A nucleic acid derivative would encode a polypeptide which retains essential biological characteristics of a natural OGR1.

"Disruption" of an OGR1 gene occurs when a fragment of genomic DNA locates and recombines with an endogenous homologous sequence. These sequence disruptions or modifications may include insertions, missense, frameshift, deletion, or substitutions, or replacements of DNA  
30 sequence, or any combination thereof. Insertions include the insertion of entire genes, which may be of animal, plant, fungal, insect, prokaryotic, or viral origin. Disruption, for example, can alter or replace a promoter, enhancer, or splice site of an OGR1 gene, and can alter the normal gene product by inhibiting its production partially or completely or by enhancing the normal gene product's activity. In a preferred embodiment, the disruption is a null disruption, wherein there is no significant  
35 expression of the OGR1 gene.



The term "native expression" refers to the expression of the full-length polypeptide encoded by the OGR1 gene, at expression levels present in the wild-type mouse. Thus, a disruption in which there is "no native expression" of the endogenous OGR1 gene refers to a partial or complete reduction of the expression of at least a portion of a polypeptide encoded by an endogenous OGR1 gene of a single cell, selected cells, or all of the cells of a mammal. The term "knockout" is a synonym for functional inactivation of the gene.

The term "construct" or "targeting construct" refers to an artificially assembled DNA segment to be transferred into a target tissue, cell line or animal. Typically, the targeting construct will include a gene or a nucleic acid sequence of particular interest, a marker gene and appropriate control sequences. As provided herein, the targeting construct of the present invention comprises an OGR1 targeting construct. An "OGR1 targeting construct" includes a DNA sequence homologous to at least one portion of an OGR1 gene and is capable of producing a disruption in an OGR1 gene in a host cell.

The term "transgenic cell" refers to a cell containing within its genome an OGR1 gene that has been disrupted, modified, altered, or replaced completely or partially by the method of gene targeting.

The term "transgenic animal" refers to an animal that contains within its genome a specific gene that has been disrupted or otherwise modified or mutated by the method of gene targeting. "Transgenic animal" includes both the heterozygous animal (*i.e.*, one defective allele and one wild-type allele) and the homozygous animal (*i.e.*, two defective alleles).

A "host cell" includes an individual cell or cell culture that can be or has been a recipient for vector(s) or for incorporation of nucleic acid molecules and/or proteins. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent due to natural, accidental, or deliberate mutation. A host cell includes cells transfected with the constructs of the present invention.

The term "modulates" or "modulation" as used herein refers to the decrease, inhibition, reduction, amelioration, increase or enhancement of OGR1 function, expression, activity, or alternatively a phenotype associated with OGR1.

The term "ameliorates" or "amelioration" as used herein refers to a decrease, reduction or elimination of a condition, disease, disorder, or phenotype, including an abnormality or symptom associated with OGR1.

The term "abnormality" refers to any disease, disorder, condition, or phenotype in which a disruption of an OGR1 gene is implicated, including pathological conditions and behavioral observations.

#### Brief Description of the Drawings

Figure 1 shows the polynucleotide sequence for a human OGR1 gene (SEQ ID NO:1).  
Figure 2 shows the amino acid sequence for a human OGR1 protein (SEQ ID NO:2).

Figures 3-4 show the location and extent of the disrupted portion of the OGR1 gene, as well as the nucleotide sequences flanking the *Neo<sup>r</sup>* insert in the targeting construct. Figure 4 shows the sequences identified as SEQ ID NO:3 and SEQ ID NO:4, which were used as the 5'- and 3'- targeting arms (including the homologous sequences) in the OGR1 targeting construct, respectively.

5        Figure 5 shows a graph comparing the total distance traveled by wild-type mice (+/+) and homozygous mutant mice (-/-) in the open field test.

Figure 6 shows a graph comparing the mean time for wild-type mice (+/+) and homozygous mutant mice (-/-) to fall from the rotarod during rotarod testing.

10       Figure 7 shows the startle response profiles of wild-type mice (+/+) and homozygous mutant mice (-/-).

Figure 8 shows a graph comparing the prepulse inhibition exhibited by homozygous mutant mice (-/-) relative to wild-type mice (+/+) during startle testing.

### Detailed Description of the Invention

15       The invention is based, in part, on the evaluation of the expression and role of genes and gene expression products, primarily those associated with an OGR1 gene. Among other uses or applications, the invention permits the definition of disease pathways and the identification of diagnostically and therapeutically useful targets. For example, genes that are mutated or down-regulated under disease conditions may be involved in causing or exacerbating the disease condition. Treatments directed at up-regulating the activity of such genes or treatments that involve alternate  
20       pathways, may ameliorate the disease condition.

### Generation of Targeting Construct

25       The targeting construct of the present invention may be produced using standard methods known in the art. (see, e.g., Sambrook, *et al.*, 1989, *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York; E.N. Glover (eds.), 1985, *DNA Cloning: A Practical Approach*, Volumes I and II; M.J. Gait (ed.), 1984, *Oligonucleotide Synthesis*; B.D. Hames & S.J. Higgins (eds.), 1985, *Nucleic Acid Hybridization*; B.D. Hames & S.J. Higgins (eds.), 1984, *Transcription and Translation*; R.I. Freshney (ed.), 1986, *Animal Cell Culture*; Immobilized Cells and Enzymes, IRL Press, 1986; B. Perbal, 1984, *A Practical Guide To Molecular Cloning*; F.M. Ausubel *et al.*, 1994, *Current Protocols in Molecular Biology*, John  
30       Wiley & Sons, Inc.). For example, the targeting construct may be prepared in accordance with conventional ways, where sequences may be synthesized, isolated from natural sources, manipulated, cloned, ligated, subjected to *in vitro* mutagenesis, primer repair, or the like. At various stages, the joined sequences may be cloned, and analyzed by restriction analysis, sequencing, or the like.

35       The targeting DNA can be constructed using techniques well known in the art. For example, the targeting DNA may be produced by chemical synthesis of oligonucleotides, nick-translation of a double-stranded DNA template, polymerase chain-reaction amplification of a sequence (or ligase

chain reaction amplification), purification of prokaryotic or target cloning vectors harboring a sequence of interest (*e.g.*, a cloned cDNA or genomic DNA, synthetic DNA or from any of the aforementioned combination) such as plasmids, phagemids, YACs, cosmids, bacteriophage DNA, other viral DNA or replication intermediates, or purified restriction fragments thereof, as well as other sources of single and double-stranded polynucleotides having a desired nucleotide sequence. Moreover, the length of homology may be selected using known methods in the art. For example, selection may be based on the sequence composition and complexity of the predetermined endogenous target DNA sequence(s).

The targeting construct of the present invention typically comprises a first sequence homologous to a portion or region of the OGR1 gene and a second sequence homologous to a second portion or region of the OGR1 gene. The targeting construct may further comprise a positive selection marker, which is preferably positioned in between the first and the second DNA sequences that are homologous to a portion or region of the target DNA sequence. The positive selection marker may be operatively linked to a promoter and a polyadenylation signal.

Other regulatory sequences known in the art may be incorporated into the targeting construct to disrupt or control expression of a particular gene in a specific cell type. In addition, the targeting construct may also include a sequence coding for a screening marker, for example, green fluorescent protein (GFP), or another modified fluorescent protein.

Although the size of the homologous sequence is not critical and can range from as few as about 15-20 base pairs to as many as 100 kb, preferably each fragment is greater than about 1 kb in length, more preferably between about 1 and about 10 kb, and even more preferably between about 1 and about 5 kb. One of skill in the art will recognize that although larger fragments may increase the number of homologous recombination events in ES cells, larger fragments will also be more difficult to clone.

In a preferred embodiment of the present invention, the targeting construct is prepared directly from a plasmid genomic library using the methods described in pending U.S. Patent Application Ser. No.: 08/971,310, filed November 17, 1997, the disclosure of which is incorporated herein in its entirety. Generally, a sequence of interest is identified and isolated from a plasmid library in a single step using, for example, long-range PCR. Following isolation of this sequence, a second polynucleotide that will disrupt the target sequence can be readily inserted between two regions encoding the sequence of interest. In accordance with this aspect, the construct is generated in two steps by (1) amplifying (for example, using long-range PCR) sequences homologous to the target sequence, and (2) inserting another polynucleotide (for example a selectable marker) into the PCR product so that it is flanked by the homologous sequences. Typically, the vector is a plasmid from a plasmid genomic library. The completed construct is also typically a circular plasmid.

In another embodiment, the targeting construct is designed in accordance with the regulated positive selection method described in U.S. Patent Application Ser. No. 09/954,483, filed September 17, 2001, the disclosure of which is incorporated herein in its entirety. The targeting construct is designed to include a PGK-*neo* fusion gene having two *lacO* sites, positioned in the PGK promoter and an NLS-*lacI* gene comprising a lac repressor fused to sequences encoding the NLS from the SV40 T antigen.

In another embodiment, the targeting construct may contain more than one selectable marker gene, including a negative selectable marker, such as the herpes simplex virus tk (HSV-tk) gene. The negative selectable marker may be operatively linked to a promoter and a polyadenylation signal. (see, e.g., U.S. Patent No. 5,464,764; U.S. Patent No. 5,487,992; U.S. Patent No. 5,627,059; and U.S. Patent No. 5,631,153).

#### Generation of Cells and Confirmation of Homologous Recombination Events

Once an appropriate targeting construct has been prepared, the targeting construct may be introduced into an appropriate host cell using any method known in the art. Various techniques may be employed in the present invention, including, for example: pronuclear microinjection; retrovirus mediated gene transfer into germ lines; gene targeting in embryonic stem cells; electroporation of embryos; sperm-mediated gene transfer; and calcium phosphate/DNA co-precipitates, microinjection of DNA into the nucleus, bacterial protoplast fusion with intact cells, transfection, polycations, e.g., polybrene, polyornithine, etc., or the like (see, e.g., U.S. Patent No. 4,873,191; Van der Putten, *et al.*, 1985, *Proc. Natl. Acad. Sci. USA* 82:6148-6152; Thompson, *et al.*, 1989, *Cell* 56:313-321; Lo, 1983, *Mol Cell. Biol.* 3:1803-1814; Lavitrano, *et al.*, 1989, *Cell*, 57:717-723). Various techniques for transforming mammalian cells are known in the art. (see, e.g., Gordon, 1989, *Intl. Rev. Cytol.*, 115:171-229; Keown *et al.*, 1989, *Methods in Enzymology*; Keown *et al.*, 1990, *Methods and Enzymology*, Vol. 185, pp. 527-537; Mansour *et al.*, 1988, *Nature*, 336:348-352).

In a preferred aspect of the present invention, the targeting construct is introduced into host cells by electroporation. In this process, electrical impulses of high field strength reversibly permeabilize biomembranes allowing the introduction of the construct. The pores created during electroporation permit the uptake of macromolecules such as DNA. (see, e.g., Potter, H., *et al.*, 1984, *Proc. Nat'l. Acad. Sci. U.S.A.* 81:7161-7165).

Any cell type capable of homologous recombination may be used in the practice of the present invention. Examples of such target cells include cells derived from vertebrates including mammals such as humans, bovine species, ovine species, murine species, simian species, and other eucaryotic organisms such as filamentous fungi, and higher multicellular organisms such as plants.

Preferred cell types include embryonic stem (ES) cells, which are typically obtained from pre-implantation embryos cultured *in vitro*. (see, e.g., Evans, M. J., *et al.*, 1981, *Nature* 292:154-156; Bradley, M. O., *et al.*, 1984, *Nature* 309:255-258; Gossler *et al.*, 1986, *Proc. Natl. Acad. Sci. USA*

83:9065-9069; and Robertson, *et al.*, 1986, *Nature* 322:445-448). The ES cells are cultured and prepared for introduction of the targeting construct using methods well known to the skilled artisan. (see, e.g., Robertson, E. J. ed. "Teratocarcinomas and Embryonic Stem Cells, a Practical Approach", IRL Press, Washington D.C., 1987; Bradley *et al.*, 1986, *Current Topics in Devel. Biol.* 20:357-371; 5 by Hogan *et al.*, in "Manipulating the Mouse Embryo": A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor N.Y., 1986; Thomas *et al.*, 1987, *Cell* 51:503; Koller *et al.*, 1991, *Proc. Natl. Acad. Sci. USA*, 88:10730; Dorin *et al.*, 1992, *Transgenic Res.* 1:101; and Veis *et al.*, 1993, *Cell* 75:229). The ES cells that will be inserted with the targeting construct are derived from an embryo or blastocyst of the same species as the developing embryo into which they are to be 10 introduced. ES cells are typically selected for their ability to integrate into the inner cell mass and contribute to the germ line of an individual when introduced into the mammal in an embryo at the blastocyst stage of development. Thus, any ES cell line having this capability is suitable for use in the practice of the present invention.

The present invention may also be used to knock out or otherwise modify or disrupt genes in 15 other cell types, such as stem cells. By way of example, stem cells may be myeloid, lymphoid, or neural progenitor and precursor cells. These cells comprising a knock out, modification or disruption of a gene may be particularly useful in the study of OGR1 gene function in individual developmental pathways. Stem cells may be derived from any vertebrate species, such as mouse, rat, dog, cat, pig, rabbit, human, non-human primates and the like.

20 After the targeting construct has been introduced into cells, the cells in which successful gene targeting has occurred are identified. Insertion of the targeting construct into the targeted gene is typically detected by identifying cells for expression of the marker gene. In a preferred embodiment, the cells transformed with the targeting construct of the present invention are subjected to treatment with an appropriate agent that selects against cells not expressing the selectable marker. Only those 25 cells expressing the selectable marker gene survive and/or grow under certain conditions. For example, cells that express the introduced neomycin resistance gene are resistant to the compound G418, while cells that do not express the neo gene marker are killed by G418. If the targeting construct also comprises a screening marker such as GFP, homologous recombination can be identified through screening cell colonies under a fluorescent light. Cells that have undergone 30 homologous recombination will have deleted the GFP gene and will not fluoresce.

If a regulated positive selection method is used in identifying homologous recombination events, the targeting construct is designed so that the expression of the selectable marker gene is regulated in a manner such that expression is inhibited following random integration but is permitted (derepressed) following homologous recombination. More particularly, the transfected cells are 35 screened for expression of the *neo* gene, which requires that (1) the cell was successfully electroporated, and (2) *lac* repressor inhibition of *neo* transcription was relieved by homologous

recombination. This method allows for the identification of transfected cells and homologous recombinants to occur in one step with the addition of a single drug.

Alternatively, a positive-negative selection technique may be used to select homologous recombinants. This technique involves a process in which a first drug is added to the cell population, for example, a neomycin-like drug to select for growth of transfected cells, *i.e.* positive selection. A second drug, such as FIAU is subsequently added to kill cells that express the negative selection marker, *i.e.* negative selection. Cells that contain and express the negative selection marker are killed by a selecting agent, whereas cells that do not contain and express the negative selection marker survive. For example, cells with non-homologous insertion of the construct express HSV thymidine kinase and therefore are sensitive to the herpes drugs such as gancyclovir (GANC) or FIAU (1-(2-deoxy 2-fluoro-B-D-arabinofluranosyl)-5-iodouracil). (*see, e.g., Mansour et al., Nature* 336:348-352: (1988); Capecchi, *Science* 244:1288-1292, (1989); Capecchi, *Trends in Genet.* 5:70-76 (1989)).

Successful recombination may be identified by analyzing the DNA of the selected cells to confirm homologous recombination. Various techniques known in the art, such as PCR and/or Southern analysis may be used to confirm homologous recombination events.

Homologous recombination may also be used to disrupt genes in stem cells, and other cell types, which are not totipotent embryonic stem cells. By way of example, stem cells may be myeloid, lymphoid, or neural progenitor and precursor cells. Such transgenic cells may be particularly useful in the study of OGR1 gene function in individual developmental pathways. Stem cells may be derived from any vertebrate species, such as mouse, rat, dog, cat, pig, rabbit, human, non-human primates and the like.

In cells that are not totipotent, it may be desirable to knock out both copies of the target using methods that are known in the art. For example, cells comprising homologous recombination at a target locus that have been selected for expression of a positive selection marker (*e.g., Neo<sup>r</sup>*) and screened for non-random integration, can be further selected for multiple copies of the selectable marker gene by exposure to elevated levels of the selective agent (*e.g., G418*). The cells are then analyzed for homozygosity at the target locus. Alternatively, a second construct can be generated with a different positive selection marker inserted between the two homologous sequences. The two constructs can be introduced into the cell either sequentially or simultaneously, followed by appropriate selection for each of the positive marker genes. The final cell is screened for homologous recombination of both alleles of the target.

#### Production of Transgenic Animals

Selected cells are then injected into a blastocyst (or other stage of development suitable for the purposes of creating a viable animal, such as, for example, a morula) of an animal (*e.g., a mouse*) to form chimeras (*see e.g., Bradley, A. in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach*, E. J. Robertson, ed., IRL, Oxford, pp. 113-152 (1987)). Alternatively, selected ES cells

can be allowed to aggregate with dissociated mouse embryo cells to form the aggregation chimera. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the embryo brought to term. Chimeric progeny harbouring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA. In one embodiment, chimeric progeny mice are used to generate a mouse with a heterozygous disruption in the OGR1 gene. Heterozygous transgenic mice can then be mated. It is well known in the art that typically  $\frac{1}{4}$  of the offspring of such matings will have a homozygous disruption in the OGR1 gene.

The heterozygous and homozygous transgenic mice can then be compared to normal, wild-type mice to determine whether disruption of the OGR1 gene causes phenotypic changes, especially pathological changes. For example, heterozygous and homozygous mice may be evaluated for phenotypic changes by physical examination, necropsy, histology, clinical chemistry, complete blood count, body weight, organ weights, and cytological evaluation of bone marrow. Phenotypic changes may also comprise behavioral modifications or abnormalities.

In one embodiment, the phenotype (or phenotypic change) associated with a disruption in the OGR1 gene is placed into or stored in a database. Preferably, the database includes: (i) genotypic data (e.g., identification of the disrupted gene) and (ii) phenotypic data (e.g., phenotype(s) resulting from the gene disruption) associated with the genotypic data. The database is preferably electronic. In addition, the database is preferably combined with a search tool so that the database is searchable.

#### Conditional Transgenic Animals

The present invention further contemplates conditional transgenic or knockout animals, such as those produced using recombination methods. Bacteriophage P1 Cre recombinase and flp recombinase from yeast plasmids are two non-limiting examples of site-specific DNA recombinase enzymes that cleave DNA at specific target sites (lox P sites for cre recombinase and frt sites for flp recombinase) and catalyze a ligation of this DNA to a second cleaved site. A large number of suitable alternative site-specific recombinases have been described, and their genes can be used in accordance with the method of the present invention. Such recombinases include the Int recombinase of bacteriophage  $\lambda$  (with or without Xis) (Weisberg, R. *et al.*, in *Lambda II*, (Hendrix, R., *et al.*, Eds.), Cold Spring Harbor Press, Cold Spring Harbor, NY, pp. 211-50 (1983), herein incorporated by reference); TpnI and the  $\beta$ -lactamase transposons (Mercier, *et al.*, *J. Bacteriol.*, 172:3745-57 (1990)); the Tn3 resolvase (Flanagan & Fennwald *J. Molec. Biol.*, 206:295-304 (1989); Stark, *et al.*, *Cell*, 58:779-90 (1989)); the yeast recombinases (Matsuzaki, *et al.*, *J. Bacteriol.*, 172:610-18 (1990)); the *B. subtilis* SpoIVC recombinase (Sato, *et al.*, *J. Bacteriol.* 172:1092-98 (1990)); the Flp recombinase (Schwartz & Sadowski, *J. Molec. Biol.*, 205:647-658 (1989); Parsons, *et al.*, *J. Biol. Chem.*, 265:4527-33 (1990); Golig & Lindquist, *Cell*, 59:499-509 (1989); Amin, *et al.*, *J. Molec. Biol.*, 214:55-72 (1990)); the Hin recombinase (Glasgow, *et al.*, *J. Biol. Chem.*, 264:10072-82 (1989));

immunoglobulin recombinases (Malynn, *et al.*, *Cell*, 54:453-460 (1988)); and the Cin recombinase (Haffter & Bickle, *EMBO J.*, 7:3991-3996 (1988); Hubner, *et al.*, *J. Molec. Biol.*, 205:493-500 (1989)), all herein incorporated by reference. Such systems are discussed by Echols (*J. Biol. Chem.* 265:14697-14700 (1990)); de Villartay (*Nature*, 335:170-74 (1988)); Craig, (*Ann. Rev. Genet.*, 22:77-105 (1988)); Poyart-Salmeron, *et al.*, (*EMBO J.* 8:2425-33 (1989)); Hunger-Bertling, *et al.*, (*Mol. Cell. Biochem.*, 92:107-16 (1990)); and Cregg & Madden (*Mol. Gen. Genet.*, 219:320-23 (1989)), all herein incorporated by reference.

Cre has been purified to homogeneity, and its reaction with the loxP site has been extensively characterized (Abremski & Hess *J. Mol. Biol.* 259:1509-14 (1984), herein incorporated by reference).

Cre protein has a molecular weight of 35,000 and can be obtained commercially from New England Nuclear/Du Pont. The cre gene (which encodes the Cre protein) has been cloned and expressed (Abremski, *et al.*, *Cell* 32:1301-11 (1983), herein incorporated by reference). The Cre protein mediates recombination between two loxP sequences (Sternberg, *et al.*, *Cold Spring Harbor Symp. Quant. Biol.* 45:297-309 (1981)), which may be present on the same or different DNA molecule.

Because the internal spacer sequence of the loxP site is asymmetrical, two loxP sites can exhibit directionality relative to one another (Hoess & Abremski *Proc. Natl. Acad. Sci. U.S.A.* 81:1026-29 (1984)). Thus, when two sites on the same DNA molecule are in a directly repeated orientation, Cre will excise the DNA between the sites (Abremski, *et al.*, *Cell* 32:1301-11 (1983)). However, if the sites are inverted with respect to each other, the DNA between them is not excised after recombination but is simply inverted. Thus, a circular DNA molecule having two loxP sites in direct orientation will recombine to produce two smaller circles, whereas circular molecules having two loxP sites in an inverted orientation simply invert the DNA sequences flanked by the loxP sites. In addition, recombinase action can result in reciprocal exchange of regions distal to the target site when targets are present on separate DNA molecules.

Recombinases have important application for characterizing gene function in knockout models. When the constructs described herein are used to disrupt OGR1 genes, a fusion transcript can be produced when insertion of the positive selection marker occurs downstream (3') of the translation initiation site of the OGR1 gene. The fusion transcript could result in some level of protein expression with unknown consequence. It has been suggested that insertion of a positive selection marker gene can affect the expression of nearby genes. These effects may make it difficult to determine gene function after a knockout event since one could not discern whether a given phenotype is associated with the inactivation of a gene, or the transcription of nearby genes. Both potential problems are solved by exploiting recombinase activity. When the positive selection marker is flanked by recombinase sites in the same orientation, the addition of the corresponding recombinase will result in the removal of the positive selection marker. In this way, effects caused by the positive selection marker or expression of fusion transcripts are avoided.



In one embodiment, purified recombinase enzyme is provided to the cell by direct microinjection. In another embodiment, recombinase is expressed from a co-transfected construct or vector in which the recombinase gene is operably linked to a functional promoter. An additional aspect of this embodiment is the use of tissue-specific or inducible recombinase constructs that allow the choice of when and where recombination occurs. One method for practicing the inducible forms of recombinase-mediated recombination involves the use of vectors that use inducible or tissue-specific promoters or other gene regulatory elements to express the desired recombinase activity. The inducible expression elements are preferably operatively positioned to allow the inducible control or activation of expression of the desired recombinase activity. Examples of such inducible promoters or other gene regulatory elements include, but are not limited to, tetracycline, metallothionine, ecdysone, and other steroid-responsive promoters, rapamycin responsive promoters, and the like (No, *et al.*, *Proc. Natl. Acad. Sci. USA*, 93:3346-51 (1996); Furth, *et al.*, *Proc. Natl. Acad. Sci. USA*, 91:9302-6 (1994)). Additional control elements that can be used include promoters requiring specific transcription factors such as viral, promoters. Vectors incorporating such promoters would only express recombinase activity in cells that express the necessary transcription factors.

#### Models for Disease

The cell- and animal-based systems described herein can be utilized as models for diseases. Animals of any species, including, but not limited to, mice, rats, rabbits, guinea pigs, pigs, micro-pigs, goats, and non-human primates, *e.g.*, baboons, monkeys, and chimpanzees may be used to generate disease animal models. In addition, cells from humans may be used. These systems may be used in a variety of applications. Such assays may be utilized as part of screening strategies designed to identify agents, such as compounds that are capable of ameliorating disease symptoms. Thus, the animal- and cell-based models may be used to identify drugs, pharmaceuticals, therapies and interventions that may be effective in treating disease.

Cell-based systems may be used to identify compounds that may act to ameliorate disease symptoms. For example, such cell systems may be exposed to a compound suspected of exhibiting an ability to ameliorate disease symptoms, at a sufficient concentration and for a time sufficient to elicit such an amelioration of disease symptoms in the exposed cells. After exposure, the cells are examined to determine whether one or more of the disease cellular phenotypes has been altered to resemble a more normal or more wild-type, non-disease phenotype.

In addition, animal-based disease systems, such as those described herein, may be used to identify compounds capable of ameliorating disease symptoms. Such animal models may be used as test substrates for the identification of drugs, pharmaceuticals, therapies, and interventions that may be effective in treating a disease or other phenotypic characteristic of the animal. For example, animal models may be exposed to a compound or agent suspected of exhibiting an ability to ameliorate disease symptoms, at a sufficient concentration and for a time sufficient to elicit such an amelioration

of disease symptoms in the exposed animals. The response of the animals to the exposure may be monitored by assessing the reversal of disorders associated with the disease. Exposure may involve treating mother animals during gestation of the model animals described herein, thereby exposing embryos or fetuses to the compound or agent that may prevent or ameliorate the disease or phenotype.

5 Neonatal, juvenile, and adult animals can also be exposed.

More particularly, using the animal models of the invention, methods of identifying agents are provided, in which such agents can be identified on the basis of their ability to affect at least one phenotype associated with a disruption in an OGR1 gene. In one embodiment, the present invention provides a method of identifying agents having an effect on OGR1 expression or function. The method includes measuring a physiological response of the animal, for example, to the agent and comparing the physiological response of such animal to a control animal, wherein the physiological response of the animal comprising a disruption in an OGR1 gene as compared to the control animal indicates the specificity of the agent. A "physiological response" is any biological or physical parameter of an animal that can be measured. Molecular assays (*e.g.*, gene transcription, protein production and degradation rates), physical parameters (*e.g.*, exercise physiology tests, measurement of various parameters of respiration, measurement of heart rate or blood pressure and measurement of bleeding time), behavioral testing, and cellular assays (*e.g.*, immunohistochemical assays of cell surface markers, or the ability of cells to aggregate or proliferate) can be used to assess a physiological response.

20 The transgenic animals and cells of the present invention may be utilized as models for diseases, disorders, or conditions associated with phenotypes relating to a disruption in an OGR1 gene.

In one aspect, the transgenic animals of the present invention exhibit hypoactivity, as described in the Examples set forth below. In a preferred embodiment, the hypoactivity is characterized by a decrease in the total distance traveled in an open field test. In accordance with this aspect, the transgenic animals may be used as an *in vivo* model for evaluating or identifying treatments for activity related disorders, such as hyperactivity or ADHD.

In another aspect, the transgenic animals of the present invention exhibit decreased motor coordination, balance or ataxia, as described in the Examples set forth below. In a preferred embodiment, the motor coordination, balance or ataxia is characterized by a decrease in the speed at which the animals fall from a rotating rod in a rotarod test. In accordance with this aspect, the transgenic animals may be used as an *in vivo* model for evaluating or identifying treatments for motor coordination related disorders, such as balance, coordination, or ataxia.

In yet another aspect, the transgenic animals of the present invention exhibit abnormal stimulus processing, as described in the Examples set forth below. In one embodiment, the abnormal stimulus processing is characterized by a decreased startle response. In another embodiment, the

abnormal stimulus processing is characterized by enhanced prepulse inhibition. Inasmuch as the enhanced prepulse inhibition is related to a deficit seen in human schizophrenic patients, the transgenic animals may be used as an *in vivo* model for evaluating or identifying treatments for schizophrenia or related disorders.

5           The present invention provides a unique animal model for testing and developing new treatments relating to the behavioral phenotypes. Analysis of the behavioral phenotype allows for the development of an animal model useful for testing, for instance, the efficacy of proposed genetic and pharmacological therapies for human genetic diseases, such as neurological, neuropsychological, or psychotic illnesses.

10           A statistical analysis of the various behaviors measured can be carried out using any conventional statistical program routinely used by those skilled in the art (such as, for example, "Analysis of Variance" or ANOVA). A "p" value of about 0.05 or less is generally considered to be statistically significant, although slightly higher p values may still be indicative of statistically  
15           significant differences. To statistically analyze abnormal behavior, a comparison is made between the behavior of a transgenic animal (or a group thereof) to the behavior of a wild-type mouse (or a group thereof), typically under certain prescribed conditions. "Abnormal behavior" as used herein refers to behavior exhibited by an animal having a disruption in the OGR1 gene, *e.g.* transgenic animal, which differs from an animal without a disruption in the OGR1 gene, *e.g.* wild-type mouse. Abnormal  
20           behavior consists of any number of standard behaviors that can be objectively measured (or observed) and compared. In the case of comparison, it is preferred that the change be statistically significant to confirm that there is indeed a meaningful behavioral difference between the knockout animal and the wild-type control animal. Examples of behaviors that may be measured or observed include, but are not limited to, ataxia, rapid limb movement, eye movement, breathing, motor activity, cognition, emotional behaviors, social behaviors, hyperactivity, hypersensitivity, anxiety, impaired learning,  
25           abnormal reward behavior, and abnormal social interaction, such as aggression.

          A series of tests may be used to measure the behavioral phenotype of the animal models of the present invention, including neurological and neuropsychological tests to identify abnormal behavior. These tests may be used to measure abnormal behavior relating to, for example, learning and memory, eating, pain, aggression, sexual reproduction, anxiety, depression, schizophrenia, and  
30           drug abuse. (*see, e.g.,* Crawley & Paylor, *Hormones and Behavior* 31:197-211 (1997)).

          The social interaction test involves exposing a mouse to other animals in a variety of settings. The social behaviors of the animals (*e.g.,* touching, climbing, sniffing, and mating) are subsequently evaluated. Differences in behaviors can then be statistically analyzed and compared (*see, e.g.,* S. E. File, *et al., Pharmacol. Bioch. Behav.* 22:941-944 (1985); R. R. Holson, *Phys. Behav.* 37:239-247  
35           (1986)). Exemplary behavioral tests include the following.

The mouse startle response test typically involves exposing the animal to a sensory (typically auditory) stimulus and measuring the startle response of the animal (*see, e.g., M. A. Geyer, et al., Brain Res. Bull.* 25:485-498 (1990); Paylor and Crawley, *Psychopharmacology* 132:169-180 (1997)). A pre-pulse inhibition test can also be used, in which the percent inhibition (from a normal startle response) is measured by "cueing" the animal first with a brief low-intensity pre-pulse prior to the startle pulse.

The electric shock test generally involves exposure to an electrified surface and measurement of subsequent behaviors such as, for example, motor activity, learning, social behaviors. The behaviors are measured and statistically analyzed using standard statistical tests. (*see, e.g., G. J. Kant, et al., Pharm. Bioch. Behav.* 20:793-797 (1984); N. J. Leidenheimer, *et al., Pharmacol. Bioch. Behav.* 30:351-355 (1988)).

The tail-pinch or immobilization test involves applying pressure to the tail of the animal and/or restraining the animal's movements. Motor activity, social behavior, and cognitive behavior are examples of the areas that are measured. (*see, e.g., M. Bertolucci D'Angic, et al., Neurochem.* 55:1208-1214 (1990)).

The novelty test generally comprises exposure to a novel environment and/or novel objects. The animal's motor behavior in the novel environment and/or around the novel object are measured and statistically analyzed. (*see, e.g., D. K. Reinstein, et al., Pharm. Bioch. Behav.* 17:193-202 (1982); B. Poucet, *Behav. Neurosci.* 103:1009-10016 (1989); R. R. Holson, *et al., Phys. Behav.* 37:231-238 (1986)). This test may be used to detect visual processing deficiencies or defects.

The learned helplessness test involves exposure to stresses, for example, noxious stimuli, which cannot be affected by the animal's behavior. The animal's behavior can be statistically analyzed using various standard statistical tests. (*see, e.g., A. Leshner, et al., Behav. Neural Biol.* 26:497-501 (1979)).

Alternatively, a tail suspension test may be used, in which the "immobile" time of the mouse is measured when suspended "upside-down" by its tail. This is a measure of whether the animal struggles, an indicator of depression. In humans, depression is believed to result from feelings of a lack of control over one's life or situation. It is believed that a depressive state can be elicited in animals by repeatedly subjecting them to aversive situations over which they have no control. A condition of "learned helplessness" is eventually reached, in which the animal will stop trying to change its circumstances and simply accept its fate. Animals that stop struggling sooner are believed to be more prone to depression. Studies have shown that the administration of certain antidepressant drugs prior to testing increases the amount of time that animals struggle before giving up.

The Morris water-maze test comprises learning spatial orientations in water and subsequently measuring the animal's behaviors, such as, for example, by counting the number of incorrect choices.

The behaviors measured are statistically analyzed using standard statistical tests. (*see, e.g., E. M. Spruijt, et al., Brain Res. 527:192-197 (1990).*)

Alternatively, a Y-shaped maze may be used (*see, e.g., McFarland, D.J., Pharmacology, Biochemistry and Behavior 32:723-726 (1989); Dellu, F., et al., Neurobiology of Learning and Memory 73:31-48 (2000).*) The Y-maze is generally believed to be a test of cognitive ability. The dimensions of each arm of the Y-maze can be, for example, approximately 40 cm x 8 cm x 20 cm, although other dimensions may be used. Each arm can also have, for example, sixteen equally spaced photobeams to automatically detect movement within the arms. At least two different tests can be performed using such a Y-maze. In a continuous Y-maze paradigm, mice are allowed to explore all three arms of a Y-maze for, *e.g., approximately 10 minutes.* The animals are continuously tracked using photobeam detection grids, and the data can be used to measure spontaneous alteration and positive bias behavior. Spontaneous alteration refers to the natural tendency of a "normal" animal to visit the least familiar arm of a maze. An alternation is scored when the animal makes two consecutive turns in the same direction, thus representing a sequence of visits to the least recently entered arm of the maze. Position bias determines egocentrically defined responses by measuring the animal's tendency to favor turning in one direction over another. Therefore, the test can detect differences in an animal's ability to navigate on the basis of allocentric or egocentric mechanisms. The two-trial Y-maze memory test measures response to novelty and spatial memory based on a free-choice exploration paradigm. During the first trial (acquisition), the animals are allowed to freely visit two arms of the Y-maze for, *e.g., approximately 15 minutes.* The third arm is blocked off during this trial. The second trial (retrieval) is performed after an intertrial interval of, *e.g., approximately 2 hours.* During the retrieval trial, the blocked arm is opened and the animal is allowed access to all three arms for, *e.g., approximately 5 minutes.* Data are collected during the retrieval trial and analyzed for the number and duration of visits to each arm. Because the three arms of the maze are virtually identical, discrimination between novelty and familiarity is dependent on "environmental" spatial cues around the room relative to the position of each arm. Changes in arm entry and duration of time spent in the novel arm in a transgenic animal model may be indicative of a role of that gene in mediating novelty and recognition processes.

The passive avoidance or shuttle box test generally involves exposure to two or more environments, one of which is noxious, providing a choice to be learned by the animal. Behavioral measures include, for example, response latency, number of correct responses, and consistency of response. (*see, e.g., R. Ader, et al., Psychon. Sci. 26:125-128 (1972); R. R. Holson, Phys. Behav. 37:221-230 (1986).*) Alternatively, a zero-maze can be used. In a zero-maze, the animals can, for example, be placed in a closed quadrant of an elevated annular platform having, *e.g., 2 open and 2 closed quadrants,* and are allowed to explore for approximately 5 minutes. This paradigm exploits an approach-avoidance conflict between normal exploratory activity and an aversion to open spaces in

rodents. This test measures anxiety levels and can be used to evaluate the effectiveness of anti-anxiolytic drugs. The time spent in open quadrants versus closed quadrants may be recorded automatically, with, for example, the placement of photobeams at each transition site.

The food avoidance test involves exposure to novel food and objectively measuring, for example, food intake and intake latency. The behaviors measured are statistically analyzed using standard statistical tests. (see, e.g., B. A. Campbell, *et al.*, *J. Comp. Physiol. Psychol.* 67:15-22 (1969)).

The elevated plus-maze test comprises exposure to a maze, without sides, on a platform, the animal's behavior is objectively measured by counting the number of maze entries and maze learning. The behavior is statistically analyzed using standard statistical tests. (see, e.g., H. A. Baldwin, *et al.*, *Brain Res. Bull.*, 20:603-606 (1988)).

The stimulant-induced hyperactivity test involves injection of stimulant drugs (e.g., amphetamines, cocaine, PCP, and the like), and objectively measuring, for example, motor activity, social interactions, cognitive behavior. The animal's behaviors are statistically analyzed using standard statistical tests. (see, e.g., P. B. S. Clarke, *et al.*, *Psychopharmacology* 96:511-520 (1988); P. Kuczenski, *et al.*, *J. Neuroscience* 11:2703-2712 (1991)).

The self-stimulation test generally comprises providing the mouse with the opportunity to regulate electrical and/or chemical stimuli to its own brain. Behavior is measured by frequency and pattern of self-stimulation. Such behaviors are statistically analyzed using standard statistical tests. (see, e.g., S. Nassif, *et al.*, *Brain Res.*, 332:247-257 (1985); W. L. Isaac, *et al.*, *Behav. Neurosci.* 103:345-355 (1989)).

The reward test involves shaping a variety of behaviors, e.g., motor, cognitive, and social, measuring, for example, rapidity and reliability of behavioral change, and statistically analyzing the behaviors measured. (see, e.g., L. E. Jarrard, *et al.*, *Exp. Brain Res.* 61:519-530 (1986)).

The DRL (differential reinforcement to low rates of responding) performance test involves exposure to intermittent reward paradigms and measuring the number of proper responses, e.g., lever pressing. Such behavior is statistically analyzed using standard statistical tests. (see, e.g., J. D. Sinden, *et al.*, *Behav. Neurosci.* 100:320-329 (1986); V. Nalwa, *et al.*, *Behav Brain Res.* 17:73-76 (1985); and A. J. Nonneman, *et al.*, *J. Comp. Physiol. Psych.* 95:588-602 (1981)).

The spatial learning test involves exposure to a complex novel environment, measuring the rapidity and extent of spatial learning, and statistically analyzing the behaviors measured. (see, e.g., N. Pitsikas, *et al.*, *Pharm. Bioch. Behav.* 38:931-934 (1991); B. Poucet, *et al.*, *Brain Res.* 37:269-280 (1990); D. Christie, *et al.*, *Brain Res.* 37:263-268 (1990); and F. Van Haaren, *et al.*, *Behav. Neurosci.* 102:481-488 (1988)). Alternatively, an open-field (of) test may be used, in which the greater distance traveled for a given amount of time is a measure of the activity level and anxiety of the animal. When the open field is a novel environment, it is believed that an approach-avoidance situation is created, in

which the animal is “torn” between the drive to explore and the drive to protect itself. Because the chamber is lighted and has no places to hide other than the corners, it is expected that a “normal” mouse will spend more time in the corners and around the periphery than it will in the center where there is no place to hide. “Normal” mice will, however, venture into the central regions as they

5 explore more and more of the chamber. It can then be extrapolated that especially anxious mice will spend most of their time in the corners, with relatively little or no exploration of the central region, whereas bold (*i.e.*, less anxious) mice will travel a greater distance, showing little preference for the periphery versus the central region.

The visual, somatosensory and auditory neglect tests generally comprise exposure to a

10 sensory stimulus, objectively measuring, for example, orientating responses, and statistically analyzing the behaviors measured. (*see, e.g.*, J. M. Vargo, *et al.*, *Exp. Neurol.* 102:199-209 (1988)).

The consummatory behavior test generally comprises feeding and drinking, and objectively measuring quantity of consumption. The behavior measured is statistically analyzed using standard statistical tests. (*see, e.g.*, P. J. Fletcher, *et al.*, *Psychopharmacol.* 102:301-308 (1990); M. G. Corda, *et*

15 *al.*, *Proc. Nat'l Acad. Sci. USA* 80:2072-2076 (1983)).

A visual discrimination test can also be used to evaluate the visual processing of an animal. One or two similar objects are placed in an open field and the animal is allowed to explore for about 5-10 minutes. The time spent exploring each object (proximity to, *i.e.*, movement within, *e.g.*, about 3-5 cm of the object is considered exploration of an object) is recorded. The animal is then removed

20 from the open field, and the objects are replaced by a similar object and a novel object. The animal is returned to the open field and the percent time spent exploring the novel object over the old object is measured (again, over about a 5-10 minute span). “Normal” animals will typically spend a higher percentage of time exploring the novel object rather than the old object. If a delay is imposed between sampling and testing, the memory task becomes more hippocampal-dependent. If no delay is

25 imposed, the task is more based on simple visual discrimination. This test can also be used for olfactory discrimination, in which the objects (preferably, simple blocks) can be sprayed or otherwise treated to hold an odor. This test can also be used to determine if the animal can make gustatory discriminations; animals that return to the previously eaten food instead of novel food exhibit gustatory neophobia.

A hot plate analgesia test can be used to evaluate an animal's sensitivity to heat or painful stimuli. For example, a mouse can be placed on an approximately 55°C hot plate and the mouse's response latency (*e.g.*, time to pick up and lick a hind paw) can be recorded. These responses are not reflexes, but rather “higher” responses requiring cortical involvement. This test may be used to

30 evaluate a nociceptive disorder.

A tail-flick test may also be used to evaluate an animal's sensitivity to heat or painful stimuli. For example, a high-intensity thermal stimulus can be directed to the tail of a mouse and the mouse's

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response latency recorded (*e.g.*, the time from onset of stimulation to a rapid flick/withdrawal from the heat source) can be recorded. These responses are simple nociceptive reflexive responses that are involuntary spinally mediated flexion reflexes. This test may also be used to evaluate a nociceptive disorder.

5           An accelerating rotarod test may be used to measure coordination and balance in mice. Animals can be, for example, placed on a rod that acts like a rotating treadmill (or rolling log). The rotarod can be made to rotate slowly at first and then progressively faster until it reaches a speed of, *e.g.*, approximately 60 rpm. The mice must continually reposition themselves in order to avoid falling off. The animals are preferably tested in at least three trials, a minimum of 20 minutes apart. Those  
10       mice that are able to stay on the rod the longest are believed to have better coordination and balance.

          A metrazol administration test can be used to screen animals for varying susceptibilities to seizures or similar events. For example, a 5mg/ml solution of metrazol can be infused through the tail vein of a mouse at a rate of, *e.g.*, approximately 0.375 ml/min. The infusion will cause all mice to experience seizures, followed by death. Those mice that enter the seizure stage the soonest are  
15       believed to be more prone to seizures. Four distinct physiological stages can be recorded: soon after the start of infusion, the mice will exhibit a noticeable "twitch", followed by a series of seizures, ending in a final tensing of the body known as "tonic extension", which is followed by death.

#### OGR1 Nucleic Acid Sequences and OGR1 Gene Products

          The present invention further contemplates use of the OGR1 gene sequence to produce OGR1  
20       gene products. OGR1 gene products may include proteins that represent functionally equivalent gene products. Such an equivalent gene product may contain deletions, additions or substitutions of amino acid residues within the amino acid sequence encoded by the gene sequences described herein, but which result in a silent change, thus producing a functionally equivalent OGR1 gene product. Amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility,  
25       hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues involved.

          For example, nonpolar (hydrophobic) amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan, and methionine; polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine, and glutamine; positively charged (basic) amino acids include arginine, lysine, and histidine; and negatively charged (acidic) amino acids include aspartic  
30       acid and glutamic acid. "Functionally equivalent", as utilized herein, refers to a protein capable of exhibiting a substantially similar *in vivo* activity as the endogenous gene products encoded by the OGR1 gene sequences. Alternatively, when utilized as part of an assay, "functionally equivalent" may refer to peptides capable of interacting with other cellular or extracellular molecules in a manner substantially similar to the way in which the corresponding portion of the endogenous gene product  
35       would.



"Percent identity" or "% identity" refers to the percentage of sequence similarity found in a comparison of two or more amino acid or nucleic acid sequences. Percent identity can be determined electronically, e.g., by using the MegAlign.TM. program (DNASTAR, Inc., Madison Wis.). The MegAlign.TM. program can create alignments between two or more sequences according to different methods, e.g., the clustal method (see, e.g., Higgins, D.G. and P.M. Sharp (1988) *Gene* 73:237-244.). The clustal algorithm groups sequences into clusters by examining the distances between all pairs. The clusters are aligned pairwise and then in groups. The percentage similarity between two amino acid sequences, e.g., sequence A and sequence B, is calculated by dividing the length of sequence A, minus the number of gap residues in sequence A, minus the number of gap residues in sequence B, into the sum of the residue matches between sequence A and sequence B, times one hundred. Gaps of low or of no similarity between the two amino acid sequences are not included in determining percentage similarity. Percent identity between nucleic acid sequences can also be counted or calculated by other methods known in the art, e.g., the Jotun Hein method (see, e.g., Hein, J. (1990) *Methods Enzymol.* 183:626-645.). Identity between sequences can also be determined by other methods known in the art, e.g., by varying hybridization conditions.

Substantially purified variants, preferably, having at least 90% sequence identity to OGR1 or to a fragment of OGR1 may be used in the methods of identifying agents that modulate OGR1 or alternatively a phenotype associated with OGR1 function as disclosed in the present invention.

Isolated and purified polynucleotides which hybridize under stringent conditions to OGR1 or a fragment of OGR1, as well as an isolated and purified OGR1 polynucleotide complementary to an OGR1 polynucleotide encoding an OGR1 amino acid sequence or a fragment thereof may be used in methods of identifying agents that modulate OGR1 or alternatively a phenotype associated with OGR1 function as disclosed by the present invention.

"Stringent conditions" refers to conditions which permit hybridization between polynucleotides and OGR1 polynucleotides. Stringent conditions can be defined by salt concentration, the concentration of organic solvent, e.g., formamide, temperature, and other conditions well known in the art. In particular, stringency can be increased by reducing the concentration of salt, increasing the concentration of formamide, or raising the hybridization temperature. For example, stringent salt concentration will ordinarily be less than about 750 mM NaCl and 75 mM trisodium citrate, preferably less than about 500 mM NaCl and 50 mM trisodium citrate, and most preferably less than about 250 mM NaCl and 25 mM trisodium citrate. Low stringency hybridization can be obtained in the absence of organic solvent, e.g., formamide, while high stringency hybridization can be obtained in the presence of at least about 35% formamide, and most preferably at least about 50% formamide. Stringent temperature conditions will ordinarily include temperatures of at least about 30°C, more preferably of at least about 37°C, and most preferably of at least about 42°C. Varying additional parameters, such as hybridization time, the

concentration of detergent, *e.g.*, sodium dodecyl sulfate (SDS), and the inclusion or exclusion of carrier DNA, are well known to those skilled in the art. Various levels of stringency are accomplished by combining these various conditions as needed. In a preferred embodiment, hybridization will occur at 30°C in 750 mM NaCl, 75 mM trisodium citrate, and 1% SDS. In a more preferred  
5 embodiment, hybridization will occur at 37°C in 500 mM NaCl, 50 mM trisodium citrate, 1% SDS, 35% formamide, and 100 µg/ml denatured salmon sperm DNA (ssDNA). In a most preferred embodiment, hybridization will occur at 42°C in 250 mM NaCl, 25 mM trisodium citrate, 1% SDS, 50% formamide, and 200 µg/ml ssDNA. Useful variations on these conditions will be readily apparent to those skilled in the art.

10 Other protein products useful according to the methods of the invention are peptides derived from or based on the OGR1 gene products produced by recombinant or synthetic means (derived peptides).

OGR1 gene products may be produced by recombinant DNA technology using techniques well known in the art. Thus, methods for preparing the gene polypeptides and peptides of the  
15 invention by expressing nucleic acids encoding gene sequences are described herein. Methods that are well known to those skilled in the art can be used to construct expression vectors containing gene protein coding sequences and appropriate transcriptional/translational control signals. These methods include, for example, *in vitro* recombinant DNA techniques, synthetic techniques and *in vivo* recombination/genetic recombination (*see, e.g.*, Sambrook et al., 1989, *supra*, and Ausubel et al.,  
20 1989, *supra*). Alternatively, RNA capable of encoding protein sequences may be chemically synthesized using, for example, automated synthesizers (*see, e.g.* Oligonucleotide Synthesis: A Practical Approach, Gait, M. J. ed., IRL Press, Oxford (1984)).

A variety of host-expression vector systems may be utilized to express the gene coding sequences of the invention. Such host-expression systems represent vehicles by which the coding  
25 sequences of interest may be produced and subsequently purified, but also represent cells that may, when transformed or transfected with the appropriate nucleotide coding sequences, exhibit the gene protein of the invention *in situ*. These include but are not limited to microorganisms such as bacteria (*e.g.*, *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing gene protein coding sequences; yeast (*e.g.* *Saccharomyces*,  
30 *Pichia*) transformed with recombinant yeast expression vectors containing the gene protein coding sequences; insect cell systems infected with recombinant virus expression vectors (*e.g.*, baculovirus) containing the gene protein coding sequences; plant cell systems infected with recombinant virus expression vectors (*e.g.*, cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (*e.g.*, Ti plasmid) containing gene protein  
35 coding sequences; or mammalian cell systems (*e.g.* COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing promoters derived from the genome of mammalian

cells (*e.g.*, metallothionine promoter) or from mammalian viruses (*e.g.*, the adenovirus late promoter; the vaccinia virus 7.5 K promoter).

In bacterial systems, a number of expression vectors may be advantageously selected depending upon the use intended for the gene protein being expressed. For example, when a large  
5 quantity of such a protein is to be produced, for the generation of antibodies or to screen peptide libraries, for example, vectors that direct the expression of high levels of fusion protein products that are readily purified may be desirable. Such vectors include, but are not limited to, the *E. coli* expression vector pUR278 (Ruther *et al.*, *EMBO J.*, 2:1791-94 (1983)), in which the gene protein coding sequence may be ligated individually into the vector in frame with the *lac Z* coding region so  
10 that a fusion protein is produced; pIN vectors (Inouye & Inouye, *Nucleic Acids Res.*, 13:3101-09 (1985); Van Heeke *et al.*, *J. Biol. Chem.*, 264:5503-9 (1989)); and the like. pGEX vectors may also be used to express foreign polypeptides as fusion proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. The pGEX vectors  
15 are designed to include thrombin or factor Xa protease cleavage sites so that the cloned OGR1 gene protein can be released from the GST moiety.

In a preferred embodiment, full length cDNA sequences are appended with in-frame Bam HI sites at the amino terminus and Eco RI sites at the carboxyl terminus using standard PCR methodologies (Innis *et al.* (eds) *PCR Protocols: A Guide to Methods and Applications*, Academic  
20 Press, San Diego (1990)) and ligated into the pGEX-2TK vector (Pharmacia, Uppsala, Sweden). The resulting cDNA construct contains a kinase recognition site at the amino terminus for radioactive labeling and glutathione S-transferase sequences at the carboxyl terminus for affinity purification (Nilsson *et al.*, *EMBO J.*, 4: 1075-80 (1985); Zabeau *et al.*, *EMBO J.*, 1: 1217-24 (1982)).

In an insect system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a  
25 vector to express foreign genes. The virus grows in *Spodoptera frugiperda* cells. The gene coding sequence may be cloned individually into non-essential regions (for example the polyhedrin gene) of the virus and placed under control of an AcNPV promoter (for example the polyhedrin promoter). Successful insertion of gene coding sequence will result in inactivation of the polyhedrin gene and production of non-occluded recombinant virus (*i.e.*, virus lacking the proteinaceous coat coded for by the polyhedrin gene). These recombinant viruses are then used to infect *Spodoptera frugiperda* cells  
30 in which the inserted gene is expressed (*see, e.g.*, Smith *et al.*, *J. Virol.* 46: 584-93 (1983); U.S. Patent No. 4,745,051).

In mammalian host cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, the gene coding sequence of interest may  
35 be ligated to an adenovirus transcription/translation control complex, *e.g.*, the late promoter and tripartite leader sequence. This chimeric gene may then be inserted in the adenovirus genome by *in*

*vitro* or *in vivo* recombination. Insertion in a non-essential region of the viral genome (*e.g.*, region E1 or E3) will result in a recombinant virus that is viable and capable of expressing gene protein in infected hosts. (*e.g.*, see Logan *et al.*, *Proc. Natl. Acad. Sci. USA*, 81:3655-59 (1984)). Specific initiation signals may also be required for efficient translation of inserted gene coding sequences.

5 These signals include the ATG initiation codon and adjacent sequences. In cases where an entire gene, including its own initiation codon and adjacent sequences, is inserted into the appropriate expression vector, no additional translational control signals may be needed. However, in cases where only a portion of the gene coding sequence is inserted, exogenous translational control signals, including, perhaps, the ATG initiation codon, must be provided. Furthermore, the initiation codon  
10 must be in phase with the reading frame of the desired coding sequence to ensure translation of the entire insert. These exogenous translational control signals and initiation codons can be of a variety of origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of appropriate transcription enhancer elements, transcription terminators, etc. (*see* Bitter *et al.*, *Methods in Enzymol.*, 153:516-44 (1987)).

15 In addition, a host cell strain may be chosen that modulates the expression of the inserted sequences, or modifies and processes the gene product in the specific fashion desired. Such modifications (*e.g.*, glycosylation) and processing (*e.g.*, cleavage) of protein products may be important for the function of the protein. Different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins. Appropriate cell lines  
20 or host systems can be chosen to ensure the correct modification and processing of the foreign protein expressed. To this end, eukaryotic host cells that possess the cellular machinery for proper processing of the primary transcript, glycosylation, and phosphorylation of the gene product may be used. Such mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, 293, 3T3, WI38, etc.

25 For long-term, high-yield production of recombinant proteins, stable expression is preferred. For example, cell lines that stably express the gene protein may be engineered. Rather than using expression vectors that contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (*e.g.*, promoter, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the  
30 introduction of the foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection and allows cells that stably integrate the plasmid into their chromosomes and grow, to form foci, which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines that express the gene protein. Such  
35 engineered cell lines may be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the gene protein.

In a preferred embodiment, timing and/or quantity of expression of the recombinant protein can be controlled using an inducible expression construct. Inducible constructs and systems for inducible expression of recombinant proteins will be well known to those skilled in the art. Examples of such inducible promoters or other gene regulatory elements include, but are not limited to, tetracycline, metallothionine, ecdysone, and other steroid-responsive promoters, rapamycin responsive promoters, and the like (No et al., *Proc. Natl. Acad. Sci. USA*, 93:3346-51 (1996); Furth et al., *Proc. Natl. Acad. Sci. USA*, 91:9302-6 (1994)). Additional control elements that can be used include promoters requiring specific transcription factors such as viral, particularly HIV, promoters. In one embodiment, a Tet inducible gene expression system is utilized (Gossen et al., *Proc. Natl. Acad. Sci. USA*, 89:5547-51 (1992); Gossen et al., *Science*, 268:1766-69 (1995)). Tet Expression Systems are based on two regulatory elements derived from the tetracycline-resistance operon of the *E. coli* Tn10 transposon - the tetracycline repressor protein (TetR) and the tetracycline operator sequence (*tetO*) to which TetR binds. Using such a system, expression of the recombinant protein is placed under the control of the *tetO* operator sequence and transfected or transformed into a host cell. In the presence of TetR, which is co-transfected into the host cell, expression of the recombinant protein is repressed due to binding of the TetR protein to the *tetO* regulatory element. High-level, regulated gene expression can then be induced in response to varying concentrations of tetracycline (Tc) or Tc derivatives such as doxycycline (Dox), which compete with *tetO* elements for binding to TetR. Constructs and materials for tet inducible gene expression are available commercially from CLONTECH Laboratories, Inc., Palo Alto, CA.

When used as a component in an assay system, the gene protein may be labeled, either directly or indirectly, to facilitate detection of a complex formed between the gene protein and a test substance. Any of a variety of suitable labeling systems may be used including but not limited to radioisotopes such as <sup>125</sup>I; enzyme labeling systems that generate a detectable calorimetric signal or light when exposed to substrate; and fluorescent labels. Where recombinant DNA technology is used to produce the gene protein for such assay systems, it may be advantageous to engineer fusion proteins that can facilitate labeling, immobilization and/or detection.

Indirect labeling involves the use of a protein, such as a labeled antibody, which specifically binds to the gene product. Such antibodies include but are not limited to polyclonal, monoclonal, chimeric, single chain, Fab fragments and fragments produced by a Fab expression library.

#### Production of Antibodies

Described herein are methods for the production of antibodies capable of specifically recognizing one or more epitopes. Such antibodies may include, but are not limited to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')<sub>2</sub> fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above. Such antibodies may be used,

for example, in the detection of an OGR1 gene in a biological sample, or, alternatively, as a method for the inhibition of abnormal OGR1 gene activity. Thus, such antibodies may be utilized as part of disease treatment methods, and/or may be used as part of diagnostic techniques whereby patients may be tested for abnormal levels of OGR1 gene proteins, or for the presence of abnormal forms of such proteins.

For the production of antibodies, various host animals may be immunized by injection with the OGR1 gene, its expression product or a portion thereof. Such host animals may include but are not limited to rabbits, mice, rats, goats and chickens, to name but a few. Various adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG (*bacille Calmette-Guerin*) and *Corynebacterium parvum*.

Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of animals immunized with an antigen, such as an OGR1 gene product, or an antigenic functional derivative thereof. For the production of polyclonal antibodies, host animals such as those described above, may be immunized by injection with gene product supplemented with adjuvants as also described above.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique that provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to the hybridoma technique of Köhler and Milstein, *Nature*, 256:495-7 (1975); and U.S. Patent No. 4,376,110), the human B-cell hybridoma technique (Kosbor, *et al.*, *Immunology Today*, 4:72 (1983); Cote, *et al.*, *Proc. Natl. Acad. Sci. USA*, 80:2026-30 (1983)), and the EBV-hybridoma technique (Cole, *et al.*, in *Monoclonal Antibodies And Cancer Therapy*, Alan R. Liss, Inc., New York, pp. 77-96 (1985)). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison, *et al.*, *Proc. Natl. Acad. Sci.*, 81:6851-6855 (1984); Takeda, *et al.*, *Nature*, 314:452-54 (1985)) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those having a variable region derived from a murine mAb and a human immunoglobulin constant region.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent No. 4,946,778; Bird, *Science* 242:423-26 (1988); Huston, *et al.*, *Proc. Natl. Acad. Sci. USA*, 85:5879-83 (1988); and Ward, *et al.*, *Nature*, 334:544-46 (1989)) can be adapted to produce gene-single chain antibodies. Single chain antibodies are typically formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments that recognize specific epitopes may be generated by known techniques. For example, such fragments include but are not limited to: the F(ab')<sub>2</sub> fragments that can be produced by pepsin digestion of the antibody molecule and the Fab fragments that can be generated by reducing the disulfide bridges of the F(ab')<sub>2</sub> fragments. Alternatively, Fab expression libraries may be constructed (Huse, *et al.*, *Science*, 246:1275-81 (1989)) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

#### Screening Methods

Various animal-derived "preparations," including cells and tissues, as well as cell-free extracts, homogenates, fractions and purified proteins, may be used to determine whether a particular agent is capable of modulating an activity of OGR1 or a phenotype associated therewith. For example, such preparations may be generated according to methods well known in the art from the tissues or organs of wild-type and knockout animals. Wild-type, but not knockout, preparations will contain endogenous OGR1, as well as the native activities, interactions and effects of OGR1. Thus, when knockout and wild-type preparations are contacted with a test agent in parallel, the ability of the test agent to modulate OGR1, or a phenotype associated therewith, can be determined. Agents capable of modulating an activity of OGR1 or a phenotype associated therewith are identified as those that modulate wild-type, but not knockout, preparations. Modulation may be detected, for example, as the ability of the agent to interact with a preparation, thereby indicating interaction with the gene product itself or a product thereof. Alternatively, the agent may affect a structural, metabolic or biochemical feature of the preparation, such as enzymatic activity of the preparation related to OGR1. An inclusive discussion of the events for which modulation by a test agent may be observed is beyond the scope of this application, but will be well known by those skilled in the art.

The present invention may be employed in a process for screening for agents such as agonists, *i.e.*, agents that bind to and activate OGR1 polypeptides, or antagonists, *i.e.*, inhibit the activity or interaction of OGR1 polypeptides with its ligand. Thus, polypeptides of the invention may also be used to assess the binding of small molecule substrates and ligands in, for example, cells, cell-free preparations, chemical libraries, and natural product mixtures as known in the art. Any methods routinely used to identify and screen for agents that can modulate receptors may be used in accordance with the present invention.

The present invention provides methods for identifying and screening for agents that modulate OGR1 expression or function. More particularly, cells that contain and express OGR1 gene

sequences may be used to screen for therapeutic agents. Such cells may include non-recombinant monocyte cell lines, such as U937 (ATCC# CRL-1593), THP-1 (ATCC# TIB-202), and P388D1 (ATCC# TIB-63); endothelial cells such as HUVEC's and bovine aortic endothelial cells (BAEC's); as well as generic mammalian cell lines such as HeLa cells and COS cells, *e.g.*, COS-7 (ATCC# CRL-1651). Further, such cells may include recombinant, transgenic cell lines. For example, the transgenic mice of the invention may be used to generate cell lines, containing one or more cell types involved in a disease, that can be used as cell culture models for that disorder. While cells, tissues, and primary cultures derived from the disease transgenic animals of the invention may be utilized, the generation of continuous cell lines is preferred. For examples of techniques that may be used to derive a continuous cell line from the transgenic animals, see Small, *et al.*, *Mol. Cell Biol.*, 5:642-48 (1985).

OGR1 gene sequences may be introduced into and overexpressed in, the genome of the cell of interest. In order to overexpress an OGR1 gene sequence, the coding portion of the OGR1 gene sequence may be ligated to a regulatory sequence that is capable of driving gene expression in the cell type of interest. Such regulatory regions will be well known to those of skill in the art, and may be utilized in the absence of undue experimentation. OGR1 gene sequences may also be disrupted or underexpressed. Cells having OGR1 gene disruptions or underexpressed OGR1 gene sequences may be used, for example, to screen for agents capable of affecting alternative pathways that compensate for any loss of function attributable to the disruption or underexpression.

*In vitro* systems may be designed to identify compounds capable of binding the OGR1 gene products. Such compounds may include, but are not limited to, peptides made of D-and/or L-configuration amino acids (in, for example, the form of random peptide libraries; (*see e.g.*, Lam, *et al.*, *Nature*, 354:82-4 (1991)), phosphopeptides (in, for example, the form of random or partially degenerate, directed phosphopeptide libraries; *see, e.g.*, Songyang, *et al.*, *Cell*, 72:767-78 (1993)), antibodies, and small organic or inorganic molecules. Compounds identified may be useful, for example, in modulating the activity of OGR1 gene proteins, preferably mutant OGR1 gene proteins; elaborating the biological function of the OGR1 gene protein; or screening for compounds that disrupt normal OGR1 gene interactions or themselves disrupt such interactions.

The principle of the assays used to identify compounds that bind to the OGR1 gene protein involves preparing a reaction mixture of the OGR1 gene protein and the test compound under conditions and for a time sufficient to allow the two components to interact and bind, thus forming a complex that can be removed and/or detected in the reaction mixture. These assays can be conducted in a variety of ways. For example, one method to conduct such an assay would involve anchoring the OGR1 gene protein or the test substance onto a solid phase and detecting target protein/test substance complexes anchored on the solid phase at the end of the reaction. In one embodiment of such a



method, the OGR1 gene protein may be anchored onto a solid surface, and the test compound, which is not anchored, may be labeled, either directly or indirectly.

In practice, microtitre plates are conveniently utilized. The anchored component may be immobilized by non-covalent or covalent attachments. Non-covalent attachment may be accomplished simply by coating the solid surface with a solution of the protein and drying. Alternatively, an immobilized antibody, preferably a monoclonal antibody, specific for the protein may be used to anchor the protein to the solid surface. The surfaces may be prepared in advance and stored.

In order to conduct the assay, the nonimmobilized component is added to the coated surface containing the anchored component. After the reaction is complete, unreacted components are removed (*e.g.*, by washing) under conditions such that any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways. Where the previously nonimmobilized component is pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the previously nonimmobilized component is not pre-labeled, an indirect label can be used to detect complexes anchored on the surface; *e.g.*, using a labeled antibody specific for the previously nonimmobilized component (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody).

Alternatively, a reaction can be conducted in a liquid phase, the reaction products separated from unreacted components, and complexes detected; *e.g.*, using an immobilized antibody specific for OGR1 gene product or the test compound to anchor any complexes formed in solution, and a labeled antibody specific for the other component of the possible complex to detect anchored complexes.

Compounds that are shown to bind to a particular OGR1 gene product through one of the methods described above can be further tested for their ability to elicit a biochemical response from the OGR1 gene protein. Agonists, antagonists and/or inhibitors of the expression product can be identified utilizing assays well known in the art.

#### Antisense, Ribozymes, and Antibodies

Other agents that may be used as therapeutics include the OGR1 gene, its expression product(s) and functional fragments thereof. Additionally, agents that reduce or inhibit mutant OGR1 gene activity may be used to ameliorate disease symptoms. Such agents include antisense, ribozyme, and triple helix molecules. Techniques for the production and use of such molecules are well known to those of skill in the art.

Anti-sense RNA and DNA molecules act to directly block the translation of mRNA by hybridizing to targeted mRNA and preventing protein translation. With respect to antisense DNA, oligodeoxyribonucleotides derived from the translation initiation site, *e.g.*, between the -10 and +10 regions of the OGR1 gene nucleotide sequence of interest, are preferred.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by an endonucleolytic cleavage. The composition of ribozyme molecules must include one or more sequences complementary to the OGR1 gene mRNA, and must include the well known catalytic sequence responsible for mRNA cleavage. For this sequence, see U.S. Patent No. 5,093,246, which is incorporated by reference herein in its entirety. As such within the scope of the invention are engineered hammerhead motif ribozyme molecules that specifically and efficiently catalyze endonucleolytic cleavage of RNA sequences encoding OGR1 gene proteins.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the molecule of interest for ribozyme cleavage sites that include the following sequences, GUA, GUU and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the OGR1 gene containing the cleavage site may be evaluated for predicted structural features, such as secondary structure, that may render the oligonucleotide sequence unsuitable. The suitability of candidate sequences may also be evaluated by testing their accessibility to hybridization with complementary oligonucleotides, using ribonuclease protection assays.

Nucleic acid molecules to be used in triple helix formation for the inhibition of transcription should be single stranded and composed of deoxyribonucleotides. The base composition of these oligonucleotides must be designed to promote triple helix formation via Hoogsteen base pairing rules, which generally require sizeable stretches of either purines or pyrimidines to be present on one strand of a duplex. Nucleotide sequences may be pyrimidine-based, which will result in TAT and CGC triplets across the three associated strands of the resulting triple helix. The pyrimidine-rich molecules provide base complementarity to a purine-rich region of a single strand of the duplex in a parallel orientation to that strand. In addition, nucleic acid molecules may be chosen that are purine-rich, for example, containing a stretch of G residues. These molecules will form a triple helix with a DNA duplex that is rich in GC pairs, in which the majority of the purine residues are located on a single strand of the targeted duplex, resulting in GGC triplets across the three strands in the triplex.

Alternatively, the potential sequences that can be targeted for triple helix formation may be increased by creating a so called "switchback" nucleic acid molecule. Switchback molecules are synthesized in an alternating 5'-3', 3'-5' manner, such that they base pair with first one strand of a duplex and then the other, eliminating the necessity for a sizeable stretch of either purines or pyrimidines to be present on one strand of a duplex.

It is possible that the antisense, ribozyme, and/or triple helix molecules described herein may reduce or inhibit the transcription (triple helix) and/or translation (antisense, ribozyme) of mRNA produced by both normal and mutant OGR1 gene alleles. In order to ensure that substantially normal

levels of OGR1 gene activity are maintained, nucleic acid molecules that encode and express OGR1 polypeptides exhibiting normal activity may be introduced into cells that do not contain sequences susceptible to whatever antisense, ribozyme, or triple helix treatments are being utilized.

Alternatively, it may be preferable to coadminister normal OGR1 protein into the cell or tissue in order to maintain the requisite level of cellular or tissue OGR1 gene activity.

Anti-sense RNA and DNA, ribozyme, and triple helix molecules of the invention may be prepared by any method known in the art for the synthesis of DNA and RNA molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides and oligoribonucleotides well known in the art such as for example solid phase phosphoramidite chemical synthesis.

Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA sequences encoding the antisense RNA molecule. Such DNA sequences may be incorporated into a wide variety of vectors that incorporate suitable RNA polymerase promoters such as the T7 or SP6 polymerase promoters. Alternatively, antisense cDNA constructs that synthesize antisense RNA constitutively or inducibly, depending on the promoter used, can be introduced stably into cell lines.

Various well-known modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but are not limited to the addition of flanking sequences of ribonucleotides or deoxyribonucleotides to the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

Antibodies that are both specific for OGR1 protein, and in particular, the mutant OGR1 protein, and interfere with its activity may be used to inhibit mutant OGR1 gene function. Such antibodies may be generated against the proteins themselves or against peptides corresponding to portions of the proteins using standard techniques known in the art and as also described herein. Such antibodies include but are not limited to polyclonal, monoclonal, Fab fragments, single chain antibodies, chimeric antibodies, antibody mimetics, etc.

In instances where the OGR1 protein is intracellular and whole antibodies are used, internalizing antibodies may be preferred. However, lipofectin liposomes may be used to deliver the antibody or a fragment of the Fab region that binds to the OGR1 gene epitope into cells. Where fragments of the antibody are used, the smallest inhibitory fragment that binds to the target or expanded target protein's binding domain is preferred. For example, peptides having an amino acid sequence corresponding to the domain of the variable region of the antibody that binds to the OGR1 protein may be used. Such peptides may be synthesized chemically or produced via recombinant DNA technology using methods well known in the art (*see, e.g.*, Creighton, *Proteins: Structures and Molecular Principles* (1984) W.H. Freeman, New York 1983, *supra*; and Sambrook, *et al.*, 1989, *supra*). Alternatively, single chain neutralizing antibodies that bind to intracellular OGR1 gene epitopes may also be administered. Such single chain antibodies may be administered, for example,

by expressing nucleotide sequences encoding single-chain antibodies within the target cell population by utilizing, for example, techniques such as those described in Marasco, *et al.*, *Proc. Natl. Acad. Sci. USA*, 90:7889-93 (1993).

RNA sequences encoding OGR1 protein may be directly administered to a patient exhibiting  
5 disease symptoms, at a concentration sufficient to produce a level of OGR1 protein such that disease symptoms are ameliorated. Patients may be treated by gene replacement therapy. One or more copies of a normal OGR1 gene, or a portion of the gene that directs the production of a normal OGR1 protein with OGR1 gene function, may be inserted into cells using vectors that include, but are not limited to adenovirus, adeno-associated virus, and retrovirus vectors, in addition to other particles that introduce  
10 DNA into cells, such as liposomes. Additionally, techniques such as those described above may be utilized for the introduction of normal OGR1 gene sequences into human cells.

Cells, preferably autologous cells, containing normal OGR1 gene expressing gene sequences may then be introduced or reintroduced into the patient at positions that allow for the amelioration of disease symptoms.

#### 15 Pharmaceutical Compositions, Effective Dosages, and Routes of Administration

The identified compounds that inhibit target mutant gene expression, synthesis and/or activity can be administered to a patient at therapeutically effective doses to treat or ameliorate the disease. A therapeutically effective dose refers to that amount of the compound sufficient to result in amelioration of symptoms of the disease.

20 Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, for determining the LD<sub>50</sub> (the dose lethal to 50% of the population) and the ED<sub>50</sub> (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD<sub>50</sub>/ED<sub>50</sub>. Compounds that exhibit large therapeutic indices are preferred.  
25 While compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of  
30 circulating concentrations that include the ED<sub>50</sub> with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC<sub>50</sub> (*i.e.*, the concentration of the test compound that  
35 achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can

be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

Pharmaceutical compositions for use in accordance with the present invention may be formulated in conventional manner using one or more physiologically acceptable carriers or excipients. Thus, the compounds and their physiologically acceptable salts and solvates may be formulated for administration by inhalation or insufflation (either through the mouth or the nose) or oral, buccal, parenteral, topical, subcutaneous, intraperitoneal, intravenous, intrapleural, intraocular, intraarterial, or rectal administration. It is also contemplated that pharmaceutical compositions may be administered with other products that potentiate the activity of the compound and optionally, may include other therapeutic ingredients.

For oral administration, the pharmaceutical compositions may take the form of, for example, tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (*e.g.*, pregelatinised maize starch, polyvinylpyrrolidone or hydroxypropyl methylcellulose); fillers (*e.g.*, lactose, microcrystalline cellulose or calcium hydrogen phosphate); lubricants (*e.g.*, magnesium stearate, talc or silica); disintegrants (*e.g.*, potato starch or sodium starch glycolate); or wetting agents (*e.g.*, sodium lauryl sulphate). The tablets may be coated by methods well known in the art. Liquid preparations for oral administration may take the form of, for example, solutions, syrups or suspensions, or they may be presented as a dry product for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (*e.g.*, sorbitol syrup, cellulose derivatives or hydrogenated edible fats); emulsifying agents (*e.g.*, lecithin or acacia); non-aqueous vehicles (*e.g.*, almond oil, oily esters, ethyl alcohol or fractionated vegetable oils); and preservatives (*e.g.*, methyl or propyl-p-hydroxybenzoates or sorbic acid). The preparations may also contain buffer salts, flavoring, coloring and sweetening agents as appropriate.

Preparations for oral administration may be suitably formulated to give controlled release of the active compound.

For buccal administration the compositions may take the form of tablets or lozenges formulated in conventional manner.

For administration by inhalation, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebuliser, with the use of a suitable propellant, *e.g.*, dichlorodifluoromethane, trichlorofluoromethane, dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of *e.g.* gelatin for use in an inhaler or insufflator may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

The compounds may be formulated for parenteral administration by injection, *e.g.*, by bolus injection or continuous infusion. Formulations for injection may be presented in unit dosage form, *e.g.*, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous vehicles, and may contain  
5   formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, *e.g.*, sterile pyrogen-free water, before use.

The compounds may also be formulated in rectal compositions such as suppositories or retention enemas, *e.g.*, containing conventional suppository bases such as cocoa butter or other  
10   glycerides. Oral ingestion is possibly the easiest method of taking any medication. Such a route of administration, is generally simple and straightforward and is frequently the least inconvenient or unpleasant route of administration from the patient's point of view. However, this involves passing the material through the stomach, which is a hostile environment for many materials, including proteins and other biologically active compositions. As the acidic, hydrolytic and proteolytic  
15   environment of the stomach has evolved efficiently to digest proteinaceous materials into amino acids and oligopeptides for subsequent anabolism, it is hardly surprising that very little or any of a wide variety of biologically active proteinaceous material, if simply taken orally, would survive its passage through the stomach to be taken up by the body in the small intestine. The result, is that many proteinaceous medicaments must be taken in through another method, such as parenterally, often by  
20   subcutaneous, intramuscular or intravenous injection.

Pharmaceutical compositions may also include various buffers (*e.g.*, Tris, acetate, phosphate), solubilizers (*e.g.*, Tween, Polysorbate), carriers such as human serum albumin, preservatives (thimerosol, benzyl alcohol) and anti-oxidants such as ascorbic acid in order to stabilize  
pharmaceutical activity. The stabilizing agent may be a detergent, such as tween-20, tween-80, NP-  
25   40 or Triton X-100. EBP may also be incorporated into particulate preparations of polymeric compounds for controlled delivery to a patient over an extended period of time. A more extensive survey of components in pharmaceutical compositions is found in Remington's Pharmaceutical Sciences, 18th ed., A. R. Gennaro, ed., Mack Publishing, Easton, Pa. (1990).

In addition to the formulations described previously, the compounds may also be formulated  
30   as a depot preparation. Such long acting formulations may be administered by implantation (for example, subcutaneously or intramuscularly) or by intramuscular injection. Thus, for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

35   The compositions may, if desired, be presented in a pack or dispenser device that may contain one or more unit dosage forms containing the active ingredient. The pack may for example comprise

metal or plastic foil, such as a blister pack. The pack or dispenser device may be accompanied by instructions for administration.

### Diagnostics

A variety of methods may be employed to diagnose disease conditions associated with the OGR1 gene. Specifically, reagents may be used, for example, for the detection of the presence of OGR1 gene mutations, or the detection of either over- or under- expression of OGR1 gene mRNA.

According to the diagnostic and prognostic method of the present invention, alteration of the wild-type OGR1 gene locus is detected. In addition, the method can be performed by detecting the wild-type OGR1 gene locus and confirming the lack of a predisposition or neoplasia. "Alteration of a

wild-type gene" encompasses all forms of mutations including deletions, insertions and point mutations in the coding and noncoding regions. Deletions may be of the entire gene or only a portion of the gene. Point mutations may result in stop codons, frameshift mutations or amino acid substitutions. Somatic mutations are those that occur only in certain tissues, *e.g.*, in tumor tissue, and are not inherited in the germline. Germline mutations can be found in any of a body's tissues and are inherited. If only a single allele is somatically mutated, an early neoplastic state may be indicated.

However, if both alleles are mutated, then a late neoplastic state may be indicated. The finding of gene mutations thus provides both diagnostic and prognostic information. An OGR1 gene allele that is not deleted (*e.g.*, that found on the sister chromosome to a chromosome carrying an OGR1 gene deletion) can be screened for other mutations, such as insertions, small deletions, and point mutations.

Mutations found in tumor tissues may be linked to decreased expression of the OGR1 gene product. However, mutations leading to non-functional gene products may also be linked to a cancerous state. Point mutational events may occur in regulatory regions, such as in the promoter of the gene, leading to loss or diminution of expression of the mRNA. Point mutations may also abolish proper RNA processing, leading to loss of expression of the OGR1 gene product, or a decrease in mRNA stability or translation efficiency.

One test available for detecting mutations in a candidate locus is to directly compare genomic target sequences from cancer patients with those from a control population. Alternatively, one could sequence messenger RNA after amplification, *e.g.*, by PCR, thereby eliminating the necessity of determining the exon structure of the candidate gene. Mutations from cancer patients falling outside the coding region of the OGR1 gene can be detected by examining the non-coding regions, such as introns and regulatory sequences near or within the OGR1 gene. An early indication that mutations in noncoding regions are important may come from Northern blot experiments that reveal messenger RNA molecules of abnormal size or abundance in cancer patients as compared to control individuals.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one specific gene nucleic acid or anti-gene antibody reagent

described herein, which may be conveniently used, *e.g.*, in clinical settings, to diagnose patients exhibiting disease symptoms or at risk for developing disease.

Any cell type or tissue, including brain, cortex, subcortical region, cerebellum, brainstem, olfactory bulb, spinal cord, eye, Harderian gland, heart, lung, liver, pancreas, kidney, spleen, thymus, lymph nodes, bone marrow, skin, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, large intestine, cecum, testis, epididymis, seminal vesicle, coagulating gland, prostate gland, ovary, uterus and white fat, in which the gene is expressed may be utilized in the diagnostics described below.

DNA or RNA from the cell type or tissue to be analyzed may easily be isolated using procedures that are well known to those in the art. Diagnostic procedures may also be performed *in situ* directly upon tissue sections (fixed and/or frozen) of patient tissue obtained from biopsies or resections, such that no nucleic acid purification is necessary. Nucleic acid reagents may be used as probes and/or primers for such *in situ* procedures (*see*, for example, Nuovo, PCR *In Situ* Hybridization: Protocols and Applications, Raven Press, N.Y. (1992)).

Gene nucleotide sequences, either RNA or DNA, may, for example, be used in hybridization or amplification assays of biological samples to detect disease-related gene structures and expression. Such assays may include, but are not limited to, Southern or Northern analyses, restriction fragment length polymorphism assays, single stranded conformational polymorphism analyses, *in situ* hybridization assays, and polymerase chain reaction analyses. Such analyses may reveal both quantitative aspects of the expression pattern of the gene, and qualitative aspects of the gene expression and/or gene composition. That is, such aspects may include, for example, point mutations, insertions, deletions, chromosomal rearrangements, and/or activation or inactivation of gene expression.

Preferred diagnostic methods for the detection of gene-specific nucleic acid molecules may involve for example, contacting and incubating nucleic acids, derived from the cell type or tissue being analyzed, with one or more labeled nucleic acid reagents under conditions favorable for the specific annealing of these reagents to their complementary sequences within the nucleic acid molecule of interest. Preferably, the lengths of these nucleic acid reagents are at least 9 to 30 nucleotides. After incubation, all non-annealed nucleic acids are removed from the nucleic acid:fingerprint molecule hybrid. The presence of nucleic acids from the fingerprint tissue that have hybridized, if any such molecules exist, is then detected. Using such a detection scheme, the nucleic acid from the tissue or cell type of interest may be immobilized, for example, to a solid support such as a membrane, or a plastic surface such as that on a microtitre plate or polystyrene beads. In this case, after incubation, non-annealed, labeled nucleic acid reagents are easily removed. Detection of the remaining, annealed, labeled nucleic acid reagents is accomplished using standard techniques well-known to those in the art.



Alternative diagnostic methods for the detection of gene-specific nucleic acid molecules may involve their amplification, *e.g.*, by PCR (the experimental embodiment set forth in Mullis U.S. Patent No. 4,683,202 (1987)), ligase chain reaction (Barany, *Proc. Natl. Acad. Sci. USA*, 88:189-93 (1991)), self sustained sequence replication (Guatelli, *et al.*, *Proc. Natl. Acad. Sci. USA*, 87:1874-78 (1990)),  
5 transcriptional amplification system (Kwoh, *et al.*, *Proc. Natl. Acad. Sci. USA*, 86:1173-77 (1989)), Q-Beta Replicase (Lizardi *et al.*, *Bio/Technology*, 6:1197 (1988)), or any other nucleic acid amplification method, followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

10 In one embodiment of such a detection scheme, a cDNA molecule is obtained from an RNA molecule of interest (*e.g.*, by reverse transcription of the RNA molecule into cDNA). Cell types or tissues from which such RNA may be isolated include any tissue in which wild-type fingerprint gene is known to be expressed, including, but not limited, to brain, cortex, subcortical region, cerebellum, brainstem, olfactory bulb, spinal cord, eye, Harderian gland, heart, lung, liver, pancreas, kidney,  
15 spleen, thymus, lymph nodes, bone marrow, skin, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, large intestine, cecum, testis, epididymis, seminal vesicle, coagulating gland, prostate gland, ovary, uterus and white fat. A sequence within the cDNA is then used as the template for a nucleic acid amplification reaction, such as a PCR amplification reaction, or the like. The nucleic acid reagents used as synthesis  
20 initiation reagents (*e.g.*, primers) in the reverse transcription and nucleic acid amplification steps of this method may be chosen from among the gene nucleic acid reagents described herein. The preferred lengths of such nucleic acid reagents are at least 15-30 nucleotides. For detection of the amplified product, the nucleic acid amplification may be performed using radioactively or non-radioactively labeled nucleotides. Alternatively, enough amplified product may be made such that the  
25 product may be visualized by standard ethidium bromide staining or by utilizing any other suitable nucleic acid staining method.

Antibodies directed against wild-type or mutant gene peptides may also be used as disease diagnostics and prognostics. Such diagnostic methods, may be used to detect abnormalities in the level of gene protein expression, or abnormalities in the structure and/or tissue, cellular, or subcellular  
30 location of fingerprint gene protein. Structural differences may include, for example, differences in the size, electronegativity, or antigenicity of the mutant fingerprint gene protein relative to the normal fingerprint gene protein.

Protein from the tissue or cell type to be analyzed may easily be detected or isolated using techniques that are well known to those of skill in the art, including but not limited to western blot  
35 analysis. For a detailed explanation of methods for carrying out western blot analysis, see Sambrook, *et al.* (1989) *supra*, at Chapter 18. The protein detection and isolation methods employed herein may

also be such as those described in Harlow and Lane, for example, (Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York (1988)).

Preferred diagnostic methods for the detection of wild-type or mutant gene peptide molecules may involve, for example, immunoassays wherein fingerprint gene peptides are detected by their interaction with an anti-fingerprint gene-specific peptide antibody.

For example, antibodies, or fragments of antibodies useful in the present invention may be used to quantitatively or qualitatively detect the presence of wild-type or mutant gene peptides. This can be accomplished, for example, by immunofluorescence techniques employing a fluorescently labeled antibody (see below) coupled with light microscopic, flow cytometric, or fluorimetric detection. Such techniques are especially preferred if the fingerprint gene peptides are expressed on the cell surface.

The antibodies (or fragments thereof) useful in the present invention may, additionally, be employed histologically, as in immunofluorescence or immunoelectron microscopy, for *in situ* detection of fingerprint gene peptides. *In situ* detection may be accomplished by removing a histological specimen from a patient, and applying thereto a labeled antibody of the present invention. The antibody (or fragment) is preferably applied by overlaying the labeled antibody (or fragment) onto a biological sample. Through the use of such a procedure, it is possible to determine not only the presence of the fingerprint gene peptides, but also their distribution in the examined tissue. Using the present invention, those of ordinary skill will readily perceive that any of a wide variety of histological methods (such as staining procedures) can be modified in order to achieve such *in situ* detection.

Immunoassays for wild-type, mutant, or expanded fingerprint gene peptides typically comprise incubating a biological sample, such as a biological fluid, a tissue extract, freshly harvested cells, or cells that have been incubated in tissue culture, in the presence of a detectably labeled antibody capable of identifying fingerprint gene peptides, and detecting the bound antibody by any of a number of techniques well known in the art.

The biological sample may be brought in contact with and immobilized onto a solid phase support or carrier such as nitrocellulose, or other solid support that is capable of immobilizing cells, cell particles or soluble proteins. The support may then be washed with suitable buffers followed by treatment with the detectably labeled gene-specific antibody. The solid phase support may then be washed with the buffer a second time to remove unbound antibody. The amount of bound label on solid support may then be detected by conventional means.

The terms "solid phase support or carrier" are intended to encompass any support capable of binding an antigen or an antibody. Well-known supports or carriers include glass, polystyrene, polypropylene, polyethylene, dextran, nylon, amylases, natural and modified celluloses, polyacrylamides, gabbros, and magnetite. The nature of the carrier can be either soluble to some

extent or insoluble for the purposes of the present invention. The support material may have virtually any possible structural configuration so long as the coupled molecule is capable of binding to an antigen or antibody. Thus, the support configuration may be spherical, as in a bead, or cylindrical, as in the inside surface of a test tube, or the external surface of a rod. Alternatively, the surface may be flat such as a sheet, test strip, etc. Preferred supports include polystyrene beads. Those skilled in the art will know many other suitable carriers for binding antibody or antigen, or will be able to ascertain the same by use of routine experimentation.

The binding activity of a given lot of anti-wild-type or -mutant fingerprint gene peptide antibody may be determined according to well known methods. Those skilled in the art will be able to determine operative and optimal assay conditions for each determination by employing routine experimentation.

One of the ways in which the gene peptide-specific antibody can be detectably labeled is by linking the same to an enzyme and using it in an enzyme immunoassay (EIA) (Voller, *Ric Clin Lab*, 8:289-98 (1978) ["The Enzyme Linked Immunosorbent Assay (ELISA)", *Diagnostic Horizons* 2:1-7, 1978, Microbiological Associates Quarterly Publication, Walkersville, Md.]; Voller, *et al.*, *J. Clin. Pathol.*, 31:507-20 (1978); Butler, *Meth. Enzymol.*, 73:482-523 (1981); Maggio (ed.), *Enzyme Immunoassay*, CRC Press, Boca Raton, Fla. (1980); Ishikawa, *et al.*, (eds.) *Enzyme Immunoassay*, Igaku-Shoin, Tokyo (1981)). The enzyme that is bound to the antibody will react with an appropriate substrate, preferably a chromogenic substrate, in such a manner as to produce a chemical moiety that can be detected, for example, by spectrophotometric, fluorimetric or by visual means. Enzymes that can be used to detectably label the antibody include, but are not limited to, malate dehydrogenase, staphylococcal nuclease, delta-5-steroid isomerase, yeast alcohol dehydrogenase, alpha-glycerophosphate dehydrogenase, triose phosphate isomerase, horseradish peroxidase, alkaline phosphatase, asparaginase, glucose oxidase, beta-galactosidase, ribonuclease, urease, catalase, glucose-6-phosphate dehydrogenase, glucoamylase and acetylcholinesterase. The detection can be accomplished by colorimetric methods that employ a chromogenic substrate for the enzyme. Detection may also be accomplished by visual comparison of the extent of enzymatic reaction of a substrate in comparison with similarly prepared standards.

Detection may also be accomplished using any of a variety of other immunoassays. For example, by radioactively labeling the antibodies or antibody fragments, it is possible to detect fingerprint gene wild-type, mutant, or expanded peptides through the use of a radioimmunoassay (RIA) (*see, e.g.*, Weintraub, B., *Principles of Radioimmunoassays*, Seventh Training Course on Radioligand Assay Techniques, The Endocrine Society, March, 1986). The radioactive isotope can be detected by such means as the use of a gamma counter or a scintillation counter or by autoradiography.

It is also possible to label the antibody with a fluorescent compound. When the fluorescently labeled antibody is exposed to light of the proper wave length, its presence can then be detected due to fluorescence. Among the most commonly used fluorescent labeling compounds are fluorescein isothiocyanate, rhodamine, phycoerythrin, phycocyanin, allophycocyanin, o-phthaldehyde and fluorescamine.

The antibody can also be detectably labeled using fluorescence emitting metals such as  $^{152}\text{Eu}$ , or others of the lanthanide series. These metals can be attached to the antibody using such metal chelating groups as diethylenetriaminepentacetic acid (DTPA) or ethylenediamine-tetraacetic acid (EDTA).

The antibody also can be detectably labeled by coupling it to a chemiluminescent compound. The presence of the chemiluminescent-tagged antibody is then determined by detecting the presence of luminescence that arises during the course of a chemical reaction. Examples of particularly useful chemiluminescent labeling compounds are luminol, isoluminol, theromatic acridinium ester, imidazole, acridinium salt and oxalate ester.

Likewise, a bioluminescent compound may be used to label the antibody of the present invention. Bioluminescence is a type of chemiluminescence found in biological systems in which a catalytic protein increases the efficiency of the chemiluminescent reaction. The presence of a bioluminescent protein is determined by detecting the presence of luminescence. Important bioluminescent compounds for purposes of labeling are luciferin, luciferase and aequorin.

Throughout this application, various publications, patents and published patent applications are referred to by an identifying citation. The disclosures of these publications, patents and published patent specifications referenced in this application are hereby incorporated by reference into the present disclosure to more fully describe the state of the art to which this invention pertains.

The following examples are intended only to illustrate the present invention and should in no way be construed as limiting the subject invention.

### Examples

#### Example 1: Generation of Mice Comprising OGR1 Gene Disruptions

To investigate the role of GPCRs, disruptions in OGR1 genes were produced by homologous recombination. Specifically, transgenic mice comprising disruptions in OGR1 genes were created.

More particularly, as shown in Figure 4, an OGR1-specific targeting construct having the ability to disrupt an OGR1 gene was created, using as the targeting arms in the construct the oligonucleotide sequences identified herein as SEQ ID NO:3 and SEQ ID NO:4.

The targeting construct was introduced into ES cells derived from the 129/OlaHsd mouse substrain to generate chimeric mice. The F1 mice were generated by breeding with C57BL/6 females.

The F2 homozygous mutant mice were produced by intercrossing F1 heterozygous males and females.

The transgenic mice comprising disruptions in OGR1 genes were analyzed for phenotypic changes and expression patterns. The phenotypes associated with a disruption in the OGR1 gene were determined.

### **Example 2: Expression Analysis by RT-PCR**

5 Total RNA was isolated from the organs or tissues from adult C57BL/6 wild-type mice. RNA was DNaseI treated, and reverse transcribed using random primers. The resulting cDNA was checked for the absence of genomic contamination using primers specific to non-transcribed genomic mouse DNA. cDNAs were balanced for concentration using HPRT primers.

10 RNA transcripts were detectable in all tissues analyzed: brain, cortex, subcortical region, cerebellum, brainstem, olfactory bulb, spinal cord, eye, Harderian gland, heart, lung, liver, pancreas, kidneys, spleen, thymus, lymph nodes, bone marrow, skin, gallbladder, urinary bladder, pituitary gland, adrenal gland, salivary gland, skeletal muscle, tongue, stomach, small intestine, large intestine, cecum, testis, epididymis, seminal vesicle, coagulating gland, prostate gland, ovary, uterus and white fat.

### **15 Example 3: Expression Analysis by LacZ Reporter Gene Analysis**

*Procedure:* In general, tissues from 7-12 week old heterozygous mutant mice were analyzed for lacZ expression. Organs from heterozygous mutant mice were frozen, sectioned (10  $\mu$ m), stained and analyzed for lacZ expression using X-Gal as a substrate for beta-galactosidase, followed by a Nuclear Fast Red counterstaining.

20 In addition, for brain, wholemount staining was performed. The dissected brain was cut longitudinally, fixed and stained using X-Gal as the substrate for beta-galactosidase. The reaction was stopped by washing the brain in PBS and then fixed in PBS-buffered formaldehyde.

Wild-type control tissues were also stained for lacZ expression to reveal any background or signals due to endogenous beta-galactosidase activity. The following tissues can show staining in the  
25 wild-type control sections and are therefore not suitable for X-gal staining: small and large intestines, stomach, vas deferens and epididymis. It has been previously reported that these organs contain high levels of endogenous beta-galactosidase activity.

LacZ (beta-galactosidase) expression was detectable in brain, spinal cord, eye, thymus, pituitary gland and testis. Most striking signals are seen in brain and spinal cord. LacZ expression  
30 was not detected in: sciatic nerve, Harderian glands, spleen, lymph nodes, bone marrow, aorta, heart, lung, liver, gallbladder, pancreas, kidney, urinary bladder, trachea, larynx, esophagus, thyroid gland, adrenal glands, salivary glands, tongue, skeletal muscle, skin and female reproductive systems.

#### Brain

35 In wholemount staining strong lacZ expression was detectable throughout the brain with striking signals in cortex, thalamus, hypothalamus, inferior colliculus, cerebellum and brainstem. On frozen sections X-Gal signals were apparent in cortex, caudate putamen, fornix, internal capsule

thalamus, hypothalamus, hippocampus and preoptic area. The most prominent expression was observed in dentate gyrus and the pyramidal cell layer. In cerebellum lacZ expression was most prominent in white matter. Further X-Gal signals were found in the granular layer, molecular layer and few Purkinje cells. In brainstem many nuclei expressed lacZ strongly.

5 Spinal cord

Most motor neurons of the gray matter expressed lacZ strongly.

Eyes

Faint lacZ expression was detectable in the inner nuclear layer and ganglion layer of the retina and in the ciliary body.

10 Thymus

Few cells in the medulla expressed lacZ weakly.

Pituitary Gland

Very few cells showed faint lacZ expression.

Male Reproductive Systems

15 Testis

Spermatogenic cells of the seminiferous tubules expressed lacZ.

**Example 4: Behavioral Analysis – Open Field Test**

Hyperactivity is a real-life problem for many children and their families. It affects mainly school-age youngsters, mostly males, although teenagers and adults also may also exhibit

20 hyperactivity disorders. Hyperactivity or related disorders include the following: attention deficit hyperactivity disorder (ADHD), attention deficit disorder (ADD), hyperactive child syndrome, minimal brain dysfunction, or hyperactivity. Current treatment for hyperactivity, particularly in children, include stimulant medications and behavior modification therapy. The practice of treating hyperactive children with stimulant medications, which affect the mind or behavior, has been widely

25 criticized by the public, and most believe that stimulant medications make children lethargic or depressed, or are otherwise detrimental to their behavior. As such, the discovery of new therapeutic targets for hyperactivity would be useful for discovering new treatments for hyperactivity. OGR1, as it may be involved in activity levels and hyperactivity, may provide one such potential target.

The Open Field test is designed to examine overall locomotion and anxiety levels in mice.

30 The open field provides a novel environment that creates an approach-avoidance conflict situation in which the animal desires to explore, yet instinctively seeks to protect itself. The chamber (open field environment) is lighted in the center and has no places to hide other than the corners. A normal mouse typically spends more time in the corners and around the periphery than it does in the center. Normal mice, however, will venture into the central regions as they explore the chamber. Anxious

35 mice spend most of their time in the corners, with almost no exploration of the center, whereas bold

mice will travel more and show less preference for the periphery versus the central regions of the chamber.

Adult wild-type male mice and heterozygous male mice were used in this experiment. Animals were group housed prior to testing. Each animal was placed gently in the center of its assigned chamber. Test sessions were ten minutes long, with the experimenter out of the sight of the animals. The activity of individual mice was recorded for the ten minute test session and monitored by photobeam breaks in the x-, y-, and z-axes. Measurements taken included total distance traveled, percent of session time spent in the central region of the test field, and average velocity during the ambulatory episodes. Increases or decreases in total distance traveled over the test time may indicate hyperactivity or hypoactivity, respectively. Alterations in the regional distribution of movement may indicate anxiety (i.e. increased anxiety if there is a decrease in the time spent in the central region).

*Results:* The transgenic mice comprising disruptions in the OGR1 gene (-/-) displayed a decrease in total distance traveled in the Open Field Test, relative to wild-type control mice (+/+), as shown in Figure 5. These results indicate that the homozygous mutant mice were hypoactive, suggesting a role for the OGR1 gene in activity, and activity related disorders, such as hyperactivity or ADHD. Therefore, the OGR1 gene may be useful in identifying potential agents that can modulate activity level, e.g. agents that can ameliorate hyperactivity.

#### **Example 5: Behavioral Analysis - Rotarod Test**

The Rotarod test is designed to measure motor coordination and balance in mice. The Rotarod may also screen for ataxia phenotypes. Ataxia is defined as the failure of muscular coordination or the irregularity of muscular action. Problems in motor coordination, balance, or ataxia may occur as a result of aging, stroke, nerve injury, or the like. A variety of disorders are associated with impaired balance or impaired motor coordination. A need exists to treat impaired balance, coordination or ataxia, or to improve balance and coordination.

Animals are placed on a smooth rod, 70mm in diameter, which acts as a rotating treadmill. The rotarod rotates slowly at first, then progressively increases in speed until it reaches a speed of 60 revolutions per minute (rpm). The mice must continually reposition themselves in order to keep from falling. Mice are motivated to stay on the rod and avoid falling. Those mice generally able to stay on the rod the longest may have better coordination and balance than those that fall off early. Light beams are used for sensing an animal's fall, and the time of the fall and the speed of the rod at the time of fall are recorded. Test chambers are fully enclosed so the animals are not able to jump out or see neighboring subjects. Most mice fall from the rotarod between 30 and 90 seconds after the test has begun.

Wild-type and homozygous mutant male mice were tested on the Rotarod. The mice were allowed to move about on their wire-cage top for 30 seconds prior to testing to ensure awareness. The mice were placed on the stationary rod, facing away from the experimenter. The rotarod was

programmed to reach 60 rpm after six minutes. The animals were tested over three trials with a 20-minute rest period between trials, after which the mice were returned to fresh cages. The data were analyzed to determine the average speed of the rotating rod at the fall time over three trials. A decrease in the speed of the rod at the time of fall compared to wild-types may indicate decreased motor coordination, impaired balance, or ataxia.

Homozygous mutant mice comprising disruptions in OGR1 genes exhibited a decrease in performance during rotarod testing. Specifically, when compared to age- and gender-matched wild-type (+/+) control mice, homozygous (-/-) mutant mice displayed a significant decrease in the mean time to fall from the rotarod, as shown in Figure 6. The decrease in mean time to fall from the accelerating rotarod may indicate a problem in motor coordination, such as balance, or suggest ataxia, indicating a role for OGR1 in motor coordination, balance, and/or ataxia.

#### **Example 6: Behavioral Analysis – Startle Test**

*Startle Response:* The startle test screens for changes in the basic fundamental nervous system or muscle-related functions. This includes changes in 1) hearing - auditory processing; 2) sensory and motor processing - related to the auditory circuit and culminating in a motor related output; and 3) motor abnormalities, including skeletal muscle or motor neuron related changes. The startle reflex is a short-latency response of the skeletal musculature elicited by a sudden auditory stimulus. The startle reflex is seen across many species, making the startle response test a useful animal model for studying abnormalities in the neural control of simple behaviors and searching for treatments and causes of those abnormalities. In rats or mice, the response is usually measured in a response chamber, which allows the measurement of the whole-body flinch elicited by the stimulus. Similar stimuli are used to test the response in humans, where a blink response is measured using electromyography of the orbicularis oculi muscle.

One component of the startle reflex test is prepulse inhibition (PPI). PPI is the reduction or gating of the startle reflex response produced by a weak prestimulus presented at a brief interval, usually between 30-500ms, before the startle eliciting stimulus. Both rats and humans have been exhibit a graded increase in PPI with increasing prepulse intensities.

Deficits in PPI are observed in human schizophrenia, a debilitating disease characterized by a constellation of distinctive and predictable symptoms, such as thought disorder, delusions, and hallucinations. Deficits in PPI have been associated with dopamine overactivity, as shown by the ability to produce a loss of PPI in rats treated with dopamine agonists, such as apomorphine. PPI can be restored in apomorphine treated rats by antipsychotics in a manner that correlates with clinical antipsychotic potency and D<sub>2</sub> receptor affinity. It is also believed that neural modulation of PPI in rats is affected by circuitry linking the hippocampus (HPC), the nucleus accumbens (NAC), the subpallidum, and the pontine reticular formation. Aside from dopaminergic involvement in PPI and sensory gating, both forebrain glutamatergic and serotonergic systems have been implicated in the



pathophysiology of schizophrenia and the action of atypical antipsychotics, and both glutamatergic and serotonergic activity are important substrates modulating PPI in rats. Non competitive NMDA glutamate receptor antagonists and serotonin receptor (particularly 5-HT<sub>1B</sub>) agonists have both been shown to reduce PPI in rats.

- 5 Genetic factors may be critical determinants of sensorimotor gating in rats. This has been supported by studies showing strain related differences in the dopaminergic modulation of PPI, as well as the production through inbreeding of strains of rats whose behavior was either apomorphine-sensitive or insensitive. Rats having a disruption of the 5-HT<sub>1B</sub> were reported to have slightly elevated basal PPI compared to wild-type controls, indicating a tonic regulation of PPI by 5-HT<sub>1B</sub>.
- 10 This conclusion was supported by research showing that a 5-HT<sub>1A/1B</sub> agonist reduced PPI in wild-type mice, but not in the 5-HT<sub>1B</sub> knockouts. The investigation of the effects on PPI of disruptions of other genes could be a valuable tool for understanding the role of particular gene products in the regulation of PPI and sensorimotor gating.

- 15 The connection between the abnormalities in sensorimotor gating in schizophrenic patients and PPI are supported by the belief that brain regions frequently implicated in the pathophysiology of the disorder, are also involved in the regulation of PPI. Abnormalities at several levels of the startle gating circuitry, including the hippocampus, nucleus accumbens, striatum, globus pallidus, and thalamus, have been noted in schizophrenic patients.

The mice were tested as follows:

- 20 *Sound Response Profile.* The mice were tested in a San Diego Instruments SR-LAB sound response chamber. Each mouse was exposed to 9 stimulus types that were repeated in pseudo-random order ten times during the course of the entire 25 minute test. The stimulus types in decibels were: p80, p90, p100, p110, p120, pp80, p120, pp90, p120, pp100, and p120; where p=40 msec pulse, pp=20 msec prepulse. The length of time between a prepulse and a pulse was 100 msec (onset to
- 25 onset). The mean Vmax of the ten repetitions for each trial type was computed for each mouse.

*Pre-Pulse Inhibition.* The % prepulse inhibition compared to p120 alone was computed for each mouse at three prepulse levels from the mean Vmax values. This was computed by determining the mean "p120", "pp80p120", "pp90p120", and "pp100p120" value for each mouse and then producing the ratios of % inhibition.

- 30 The startle responses of the homozygous and wild-type mice is shown in Figure 7. Homozygous mutant mice displayed decreased startle responses. Specifically, the startle responses to sound stimuli at 110 decibels and 120 decibels were significantly decreased in homozygous mutant mice (-/-), relative to wild-type mice (+/+). Further, homozygous mutant mice displayed a trend toward a decreased startle response at 100 decibels. This decrease in startle response may be
- 35 indicative of enhanced motor processing, a motor deficit, or a decreased level of anxiety.

Homozygous mice additionally exhibited enhanced prepulse inhibition, which is opposite of a behavior seen in schizophrenic patients. Specifically, as shown in Figure 8, when a 120 decibel startle stimulus was preceded by an 85 decibel prepulse, the homozygous mutants (-/-) displayed enhanced prepulse inhibition, relative to wild-type mice (+/+). Therefore, it is possible that the OGR1 gene may provide a target for discovering therapeutics for the treatment of schizophrenia or related disorders. For example, agents that serve to decrease expression of the OGR1 gene or modulate the function of the gene or protein may be useful as anti-schizophrenic drugs.

#### **Example 7: Physical Examination**

A complete physical examination was performed on each mouse. Mice were first observed in their home cages for a number of general characteristics including activity level, behavior toward siblings, posture, grooming, breathing pattern and sounds, and movement. General body condition and size were noted as well identifying characteristics including coat color, belly color, and eye color. Following a visual inspection of the mouse in the cage, the mouse was handled for a detailed, stepwise examination. The head was examined first, including eyes, ears, and nose, noting any discharge, malformations, or other abnormalities. Lymph nodes and glands of the head and neck were palpated. Skin, hair coat, axial and appendicular skeleton, and abdomen were also examined. The limbs and torso were examined visually and palpated for masses, malformations or other abnormalities. The anogenital region was examined for discharges, staining of hair, or other changes. If the mouse defecates during the examination, the feces were assessed for color and consistency. Abnormal behavior, movement, or physical changes may indicate abnormalities in general health, growth, metabolism, motor reflexes, sensory systems, or development of the central nervous system.

#### **Example 8: Necropsy Analysis**

Necropsy was performed on mice following deep general anesthesia, cardiac puncture for terminal blood collection, and euthanasia. Body lengths and body weights were recorded for each mouse. The necropsy included detailed examination of the whole mouse, the skinned carcass, skeleton, and all major organ systems. Lesions in organs and tissues were noted during the examination. Designated organs, from which extraneous fat and connective tissue have been removed, were weighed on a balance, and the weights were recorded. Weights were obtained for the following organs: heart, liver, spleen, thymus, kidneys, and testes/epididymides.

#### **Example 9: Histopathological Analysis**

Harvested organs were fixed in about 10% neutral buffered formalin for a minimum of about 48 hours at room temperature. Tissues were trimmed and samples taken to include the major features of each organ. If any abnormalities were noted at necropsy or at the time of tissue trimming, additional sample(s), if necessary, were taken to include the abnormalities so that it is available for microscopic analysis. Tissues were placed together, according to predetermined groupings, in tissue

processing cassettes. All bones (and any calcified tissues) were decalcified with a formic acid or EDTA-based solution prior to trimming.

The infiltration of the tissues by paraffin was performed using an automated tissue processor. Steps in the cycle included dehydration through a graded series of ethanols, clearing using xylene or xylene substitute and infiltration with paraffin. Tissues were embedded in paraffin blocks with a standard orientation of specified tissues within each block. Sections were cut from each block at a thickness of about 3-5 $\mu$ m and mounted onto glass slides. After drying, the slides were stained with hematoxylin and eosin (H&E) and a glass coverslip was mounted over the sections for examination.

#### **Example 10: Hematological Analysis**

Blood samples were collected via a terminal cardiac puncture in a syringe. About one hundred microliters of each whole blood sample were transferred into tubes pre-filled with EDTA. Approximately 25 microliters of the blood was placed onto a glass slide to prepare a peripheral blood smear. The blood smears were later stained with Wright's Stain that differentially stained white blood cell nuclei, granules and cytoplasm, and allowed the identification of different cell types. The slides were analyzed microscopically by counting and noting each cell type in a total of 100 white blood cells. The percentage of each of the cell types counted was then calculated. Red blood cell morphology was also evaluated.

Microscopic examinations of blood smears were performed to provide accurate differential blood leukocyte counts. The leukocyte differential counts were provided as the percentage composition of each cell type in the blood.

#### **Example 11: Serum Chemistry**

Blood samples were collected via a terminal cardiac puncture in a syringe. One hundred microliters of each whole blood sample was transferred into a tube pre-filled with EDTA. The remainder of the blood sample was converted to serum by centrifugation in a serum tube with a gel separator. Each serum sample was then analyzed as described below. Non-terminal blood samples for aged mice are collected via retro-orbital venous puncture in capillary tubes. This procedure yields approximately 200 $\mu$ L of whole blood that is either transferred into a serum tube with a gel separator for serum chemistry analysis (see below), or into a tube pre-filled with EDTA for hematology analysis.

The serum was analyzed for the following parameters: alanine aminotransferase, albumin, alkaline phosphatase, aspartate transferase, bicarbonate, total bilirubin, blood urea nitrogen, calcium, chloride, cholesterol, creatine kinase, creatinine, globulin, glucose, high density lipoproteins (HDL), lactate dehydrogenase, low density lipoproteins (LDL), osmolality, phosphorus, potassium, total protein, sodium, and triglycerides.

**Example 12: Densitometric Analysis**

Mice were euthanized and analyzed using a PIXImus<sup>TM</sup> densitometer. An x-ray source exposed the mice to a beam of both high and low energy x-rays. The ratio of attenuation of the high and low energies allowed the separation of bone from soft tissue, and, from within the tissue samples, lean and fat. Densitometric data including Bone Mineral Density (BMD presented as g/cm<sup>2</sup>), Bone Mineral Content (BMC in g), bone and tissue area, total tissue mass, and fat as a percent of body soft tissue (presented as fat %) were obtained and recorded.

**Example 13: Embryonic Development**

Animals are genotyped using one of two methods. The first method uses the polymerase chain reaction (PCR) with target-specific and Neo primers to amplify DNA from the targeted gene. The second method uses PCR and Neo primers to "count" the number of Neo genes present per genome.

If homozygous mutant mice are not identified at weaning (3-4 weeks old), animals were assessed for lethality linked with the introduced mutation. This evaluation included embryonic, perinatal or juvenile death.

Newborn mice were genotyped 24-48 hours after birth and monitored closely for any signs of stress. Dead/dying pups were recorded and grossly inspected and if possible, genotyped. In the case of perinatal death, late gestation embryos (~E19.5, i.e., 19.5 days post-coitum) or newborn pups were analyzed, genotyped and subject to further characterization.

If there was no evidence of perinatal or juvenile lethality, heterozygous mutant mice were set up for timed pregnancies. Routinely, E10.5 embryos are analyzed for gross abnormalities and genotyped. Depending on these findings, earlier (routinely >E8.5) or later embryonic stages are characterized to identify the approximate time of death. If no homozygous mutant progeny are detected, blastocysts (E3.5) are isolated, genotyped directly or grown for 6 days in culture and then genotyped. Any suspected genotype-related gross abnormalities are recorded.

**Example 14: Fertility**

The reproductive traits of male and female homozygous mutant mice are tested to identify potential defects in spermatogenesis, oogenesis, maternal ability to support pre- or post-embryonic development, or mammary gland defects and ability of the female knockout mice to nurse their pups.

Homozygous mutant (-/-) mice of each gender were set up in a fertility mating with either a wild-type (+/+) mate or a homozygous mutant mouse of the opposite gender at about seven to about ten weeks of age. The numbers of pups born from one to three litters were recorded at birth. Three weeks later, the live pups were counted and weaned.

Males and females were separated after they had produced two litters or at six months (26 weeks) of age, whichever comes first.

**Example 15: Behavioral Analysis - Hot Plate Test**

The hot plate analgesia test was designed to indicate an animal's sensitivity to a painful stimulus. The mice were placed on a hot plate of about 55.5°C, one at a time, and latency of the mice to pick up and lick or fan a hindpaw was recorded. A built-in timer was started as soon as the subjects  
5 were placed on the hot plate surface. The timer was stopped the instant the animal lifted its paw from the plate, reacting to the discomfort. Animal reaction time was a measurement of the animal's resistance to pain. The time points to hindpaw licking or fanning, up to a maximum of about 60-seconds, was recorded. Once the behavior was observed, the animal was immediately removed from the hot plate to prevent discomfort or injury.

**10 Example 16: Behavioral Analysis – Tail Flick Test**

The tail-flick test is a test of acute nociception in which a high-intensity thermal stimulus is directed to the tail of the mouse. The time from onset of stimulation to a rapid flick/withdrawal from the heat source is recorded. This test produces a simple nociceptive reflex response that is an involuntary spinally mediated flexion reflex.

**15 Example 17: Behavioral Analysis – Metrazol Test**

To screen for phenotypes involving changes in seizure susceptibility, the Metrazol Test was be used. About 5mg/ml of Metrazol was infused through the tail vein of the mouse at a constant rate of about 0.375ml/min. The infusion caused all mice to experience seizures. Those mice entering the seizure stage the quickest were thought to be more prone to seizures in general.

20 The Metrazol test can also be used to screen for phenotypes related to epilepsy. Seven to ten adult wild-type and homozygote males were used. A fresh solution of about 5mg/ml pentylenetetrazole in approximately 0.9% NaCl was prepared prior to testing. Mice were weighed and loosely held in a restrainer. After exposure to a heat lamp to dilate the tail vein, mice were continuously infused with the pentylenetetrazole solution using a syringe pump set at a constant flow  
25 rate. The following stages were recorded: first twitch (sometimes accompanied by a squeak), beginning of the tonic/clonic seizure, tonic extension and survival time. The dose required for each phase was determined and the latency to each phase was determined between genotypes. Alterations in any stage may indicate an overall imbalance in excitatory or inhibitory neurotransmitter levels.

**Example 18: Behavioral Analysis – Tail Suspension Test**

30 The tail suspension test is a single-trial test that measures a mouse's propensity towards depression. This method for testing antidepressants in mice was reported by Steru *et al.*, (1985, *Psychopharmacology* 85(3):367-370) and is widely used as a test for a range of compounds including SSRI's, benzodiazepines, typical and atypical antipsychotics. It is believed that a depressive state can be elicited in laboratory animals by continuously subjecting them to aversive situations over which  
35 they have no control. It is reported that a condition of "learned helplessness" is eventually reached.

Mice were suspended on a metal hanger by the tail in an acoustically and visually isolated setting. Total immobility time during the six-minute test period was determined using a computer algorithm based upon measuring the force exerted by the mouse on the metal hanger. An increase in immobility time for mutant mice compared to wild-type mice may indicate increased "depression."

- 5 Animals that ceased struggling sooner may be more prone to depression. Studies have shown that the administration of antidepressants prior to testing increases the amount of time that animals struggle

As is apparent to one of skill in the art, various modifications of the above embodiments can be made without departing from the spirit and scope of this invention. These modifications and variations are within the scope of this invention.

## CLAIMS

We claim:

1. A transgenic mouse comprising a disruption in an OGR1 gene.
2. A transgenic mouse comprising a disruption in an OGR1 gene, wherein there is no native expression of endogenous OGR1 gene.
3. The transgenic mouse of claim 2, wherein the disruption is heterozygous.
4. The transgenic mouse of claim 2, wherein the disruption is homozygous.
5. The transgenic mouse of claim 4, wherein the transgenic mouse exhibits hypoactivity.
6. The transgenic mouse of claim 5, wherein the hypoactivity is characterized by a decrease in total distance traveled in an open field test.
7. The transgenic mouse of claim 6, wherein the total distance traveled is consistent with a symptom associated with human hyperactivity.
8. The transgenic mouse of claim 4, wherein the transgenic mouse exhibits a motor abnormality.
9. The transgenic mouse of claim 8, wherein the motor abnormality is impaired balance, impaired coordination, or ataxia.
10. The transgenic mouse of claim 8, wherein the motor abnormality is characterized by a decrease in fall speed in a rotarod test.
11. The transgenic mouse of claim 4, wherein the transgenic mouse exhibits abnormal stimulus processing.
12. The transgenic mouse of claim 11, wherein the abnormal stimulus processing is characterized by a decreased startle response.
13. The transgenic mouse of claim 11, wherein the abnormal stimulus processing is characterized by enhanced prepulse inhibition.
14. The transgenic mouse of claim 13, wherein the prepulse inhibition is consistent with a symptom associated with human schizophrenia.
15. A method of producing a transgenic mouse comprising a disruption in an OGR1 gene, the method comprising:
  - (a) providing a murine stem cell comprising a disruption in an OGR1 gene; and
  - (b) introducing the murine stem cell into a pseudopregnant mouse, wherein the pseudopregnant mouse gives birth to a transgenic mouse.
16. The transgenic mouse produced by the method of claim 15.
17. A targeting construct comprising:
  - (a) a first polynucleotide sequence homologous to at least a first portion of an OGR1 gene;
  - (b) a second polynucleotide sequence homologous to at least a second portion of an OGR1 gene; and

(c) a selectable marker.

18. A cell comprising a disruption in an OGR1 gene, the disruption produced using the targeting construct of claim 17.
19. A cell derived from the transgenic mouse of claim 2.
20. A cell comprising a disruption in an OGR1 gene.
21. The cell of claim 20, wherein the cell is a stem cell.
22. The cell of claim 21, wherein the stem cell is an embryonic stem cell.
23. The cell of claim 22, wherein the embryonic stem cell is a murine cell.
24. A method of identifying an agent that modulates a phenotype selected from the group consisting of hypoactivity, impaired balance, impaired motor coordination, ataxia, decreased startle response, and enhanced prepulse inhibition, the method comprising:
  - (a) contacting a test agent with OGR1; and
  - (b) determining whether the agent modulates OGR1.
25. A method of identifying an agent that modulates a phenotype selected from the group consisting of hypoactivity, impaired balance, impaired motor coordination, ataxia, decreased startle response, and enhanced prepulse inhibition, the method comprising:
  - (a) administering a test agent to an animal exhibiting a phenotype selected from the group consisting of hypoactivity, impaired balance, impaired motor coordination, ataxia, decreased startle response, and enhanced prepulse inhibition; and
  - (b) determining whether the agent modulates the phenotype.
26. A method of identifying a potential therapeutic agent for the treatment of schizophrenia, the method comprising:
  - (a) administering the potential therapeutic agent to a transgenic mouse comprising a disruption in an OGR1 gene; and
  - (b) determining whether the potential therapeutic agent modulates schizophrenia, wherein modulation of schizophrenia identifies a potential therapeutic agent for the treatment of schizophrenia.
27. A method of identifying a potential therapeutic agent for the treatment of hyperactivity or impaired balance, the method comprising:
  - (a) administering the potential therapeutic agent to a transgenic mouse comprising a disruption in an OGR1 gene; and
  - (b) determining whether the potential therapeutic agent modulates hyperactivity or balance, wherein modulation of hyperactivity or balance identifies a potential therapeutic agent for the treatment of hyperactivity or impaired balance.



28. A method of identifying a potential therapeutic agent for the treatment of schizophrenia, the method comprising:
- (a) contacting the potential therapeutic agent with OGR1;
  - (b) determining whether the agent modulates OGR1, wherein modulation of OGR1 identifies a potential therapeutic agent for the treatment of schizophrenia.
29. A method of identifying a potential therapeutic agent for the treatment of hyperactivity or impaired balance, the method comprising:
- (a) contacting the potential therapeutic agent with OGR1;
  - (b) determining whether the agent modulates OGR1, wherein modulation of OGR1 identifies a potential therapeutic agent for the treatment of hyperactivity or impaired balance.
30. A method of evaluating a potential therapeutic agent capable of affecting a condition or phenotype associated with OGR1, the method comprising:
- (a) administering the potential therapeutic agent to a transgenic mouse comprising a disruption in an OGR1 gene; and
  - (b) evaluating the effects of the agent on the transgenic mouse.
31. A method of evaluating a potential therapeutic agent capable of affecting a condition or phenotype associated with OGR1, the method comprising:
- (a) contacting the potential therapeutic agent with OGR1;
  - (b) evaluating the effects of the agent on the OGR1.
32. A method of determining whether an agent modulates OGR1, the method comprising:
- (a) providing a first preparation derived from the mouse of claim 2;
  - (b) providing a second preparation derived from a wild-type mouse;
  - (c) contacting a test agent with the first and second preparations; and
  - (d) determining whether the agent modulates the first and second preparations, wherein modulation of the second preparation but not the first preparation indicates that the agent modulates the OGR1.
33. A therapeutic agent for treating schizophrenia, wherein the agent modulates OGR1.
34. A therapeutic agent for treating schizophrenia, wherein the agent is an antagonist of OGR1.
35. A therapeutic agent for treating hyperactivity, wherein the agent modulates OGR1.
36. A therapeutic agent for treating hyperactivity, wherein the agent is an antagonist of OGR1.
37. A therapeutic agent for improving balance, wherein the agent modulates OGR1.
38. A pharmaceutical composition comprising OGR1.
39. A method of preparing a pharmaceutical composition for a condition associated with a function of OGR1, the method comprising:
- (a) identifying a compound that modulates OGR1;

- (b) synthesizing the identified compound; and
- (c) incorporating the compound into a pharmaceutical carrier.

40. A method of treating schizophrenia the method comprising administering to a subject in need a therapeutically effective amount of an agent that modulates OGR1.
41. A method of treating hyperactivity the method comprising administering to a subject in need a therapeutically effective amount of an agent that modulates OGR1.
42. A method of treating impaired balance or improving balance, the method comprising administering to a subject in need a therapeutically effective amount of an agent that modulates OGR1.
43. Phenotypic data associated with a transgenic mouse comprising a disruption in an OGR1 gene, wherein the phenotypic data is in an electronic database.

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ACTCCCAAAGTGCTGGGCTTACAGGTGTAAGCCATCATGTCCAGCCGTTAGATATTCTAGTTGAATTGGAG  
 TTGGTGGGCTAGTACACCTTCTAAATTAAATGAGTAAAGGATTTAGAATGGTGCCTGACACACAGTAGGTGC  
 TACATTTCATGTTAGTACTATTATAAACCTTTCTGCTCTGACTTTCAGGGTCTTGCCACCACCAGCGAT  
 GCCCAGCCCTTGGTAGAGCTTGAACCACCTTCTATAAACAGGATGGCGGTGGAGAGACAGGCCAGTCCCTG  
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 GCAACCTGACGGTGGCCGACCTCTTCTACATCTGCTCGCTGCCCTTCTGGCTGCAGTACGTGCTGCAGCAG  
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 GGACCTTGAAGCGGCCGTCGGCGTCAGCGTGGTTCATCTGGGCCAAGGAGCTGCTGACCAGCATCTACTTCC  
 TGATGCACGAGGAGGTCATCGAGGACGAGAACCAGCACCGCGTGTGCTTTGAGCACTACCCCATCCAGGCAT  
 GGCAGCGCGCCATCAACTACTACCGCTTCTGGTGGGCTTCTCTTCCCATCTGCCTGCTGCTGGCGTCTCT  
 ACCAGGGCATCTTGC CGCGCCGTGCGCCGAGCCACGGCACCCAGAAGAGCCGCAAGGACCAGATCCAGCGGC  
 TGGTGTCTAGCACCGTGGTTCATCTTCTGGCTGTCTCTGCCCTACCACGTGTTGCTGCTGGTGCGCAGCG  
 TCTGGGAGGCCAGCTGCGACTTCGCCAAGGGCGTTTCAACGCCTACCACTTCTCCCTCCTGCTCACCAGCT  
 TCAACTGCGTCTGCCGACCCCGTGTCTACTGCTTCGTCAGCGAGACCACCCACCGGGACCTGGCCCGCTCC  
 GCGGGGCTTCCCTGGCTTCTCTACCTGCTCCAGGACCGGCCGGCCAGGGAGGCCTACCCGCTGGGTGCCC  
 CCGAGGCTTCCGGGAAAAGCGGGGCCAGGGTGAGGAGCCCGAGCTGTTGACCAAGCTCCACCCGGCTTCC  
 AGACCCCTAACTCGCCAGGTCGGGCGGGTCCCCACGGGCAGGTTGGCTAGCCTGGGTCTCCGCGGGTG  
 GCTCCACGTGAGGCTGAGCCTTACGCCCACGGGCCACAGGCTGCGGCTCTGCTTCCCTCGCTGCGGA  
 GGCAGGGAAGCCCTGTAACTCGGAAGCCTGCTCTCGCTTGTGAGCCCGCTGGGACCGCCGAGGGTGGGA  
 ATAAGCCCCGTTGGCTCGTGGGAATAAGCCGTGCTCTGCGCGGCTGCGATGTGGCCACGCTGGGGCTG  
 CTGGTCGGGGAAAACAGTGAAGTGCCTCCCTGGCTGCT (SEQ ID NO:1)

## FIGURE 1

MGNITADNSSMSCTIDHTIHQTLAPVVYVTVLVVGF PANCLSLYFGYLQIKARNELGVYL  
 CNLTVADLFYICSLPFWLQYVLQHDNWSHGDLSQVCGILLYENIYISVGLCCISVDY  
 LAVAHPPFRFHQFRTLKAAVGVSIVIWAKELLTSIYFLMHEEVIEDENQHRVCFEHYPIQA  
 WQRAINYYRFLVGFLEFPICLLLASYGILRAVRRSHGTQKSRKDQIQRLVLSTVVIFLAC  
 FLPHYVLLLVRVSWEASCDFAKGVFNAYHFSLLLTSFNVCVADPVLVYCFVSETTHRDLARL  
 RGACLAFLTCSRTGRAREAYPLGAPEASGKSGAQGEPELLTKLHPAFQTPNSPGSGGFP  
 TGRLA (SEQ ID NO:2)

## FIGURE 2

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underlined = deleted in targeting construct

**bold** = sequence flanking Neo insert in targeting construct

ACTCCCAAAGTGCTGGGCTTACAGGTGTAAGCCATCATGTCCAGCCGTTTCAGATATTCTA  
 GTTGAATTGGAGTTGGTGGGCTAGTACACCTTCTAAATTAAATGAGTAAAGGATTTAGAA  
 TGGTGCCGTGACACACAGTAGGTGCTACATTCATGTTAGCTACTATTATAAACCTTTCCCTG  
 CCTCTGACTTTCAGGGTCTTGGCCACCACCAGCGATGCCAGCCCTTGGTAGAGCTTGAA  
**CCACCTTCTATAAACAGGATGGCGGTGGAGAGACAGGCCAGTCCCTGAGCCCATGAGGA**  
**GTTGTGGCCCCCTTCAGGCCCAAAGATGGGGAACATCACTGCAGACAACTCCTCGATGAGCT**  
**GTACCATCGACCATAACCATCCACCAGACGCTGGCCCCGGTGGTCTATGTTACCGTGCTGG**  
TGGTGGGCTTCCCGGCCAACTGCCGTGCCCTCTACTTCGGCTACCTGCAGATCAAGGCC  
GGAACGAGCTGGGCGGTACCTGTGCAACCTGACGGTGGCCGACCTCTTCTACATCTGCT  
CGCTGCCCTTCTGGCTGCAGTACGTGCTGCAGCACGACAACCTGGTCTCACGGCGACCTGT  
CCTGCCAGGTGTGCGGCATCCTCCTGTACGAGAACATCTACATCAGCGTGGGCTTCCCTCT  
GCTGCATCTCCGTGGACCGCTACCTGGCTGTGGCCCATCCCTTCCGCTTCCACCAGTTCC  
GGACCTGAAGGCGGCCGTGCGCGTCAGCGTGGTCATCTGGGCCAAGGAGCTGCTGACCA  
GCATCTACTTCCGTATGCACGAGGAGGTCACTGAGGACGAGAACCAGCACCGCGTGTGCT  
TTGAGCACTACCCCATCCAGGCATGGCAGCGCGCCATCAACTACTACCGCTTCCCTGGTGG  
GCTTCCCTCTTCCCCATCTGCCTGCTGCTGGCGTCTTACCAGGGCATCCTGCGCGCCGTGC  
GCCGGAGCCACGGCACCCAGAAGAGCCGCAAGGACCAGATCCAGCGGCTGGTGTCTAGCA  
CCGTGGTCACTTCCCTGGCCTGCTTCCCTGCCCTACCACGTGTTGCTGCTGGTGGCGAGCG  
TCTGGGAGGCCAGCTGCGACTTCGCCAAGGGCGTTTTCAACGCCTACCACTTCTCCCTCC  
TGCTCACCAGCTTCAACTGCGTCGCGACCCCGTGCTCTACTGCTTCGTCTAGCGAGACCA  
CCCACCGGGACCTGGCCCGCCTCCGCGGGGCGCTGCCTGGCCTTCCCTCACCTGCTCCAGGA  
CCGCGCCGGGCCAGGGAGCCCTACCCGCTGGGTGCCCCCGAGGCCTCCGGGAAAAGCGGGG  
CCCAGGGTGAGGAGCCCCGAGCTGTTGACCAAGCTCCACCCGGCCTTCCAGACCCCTAACT  
CGCCAGGGTCGGGCGGGTTCCCCACGGGCAGGTTGGCCTAGCCTGGGTCTCCGCGGGTG  
GCTCCACGTGAGGCCTGAGCCTTCAGCCACGGGCCTCAGGGCCTGCCGCCTCCTGCTTC  
CCTCGCTGCGGAGGCAGGGAAGCCCTGTAACCTCCGGAAGCCTGCTCTCGCTTGCTGAGC  
CCGCTGGGACCGCCGAGGGTGGGAATAAGCCCCGGTTGGCTCGTGGAATAAGCCGTGTC  
CTCTGCCGCGGCTGCGATGTGGCCACGCTGGGGCTGCTGGTGGGGGAAAACAGTGAAC  
GCGTCCCCCTGGCCTGCT (SEQ ID NO:1)

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FIGURE 3

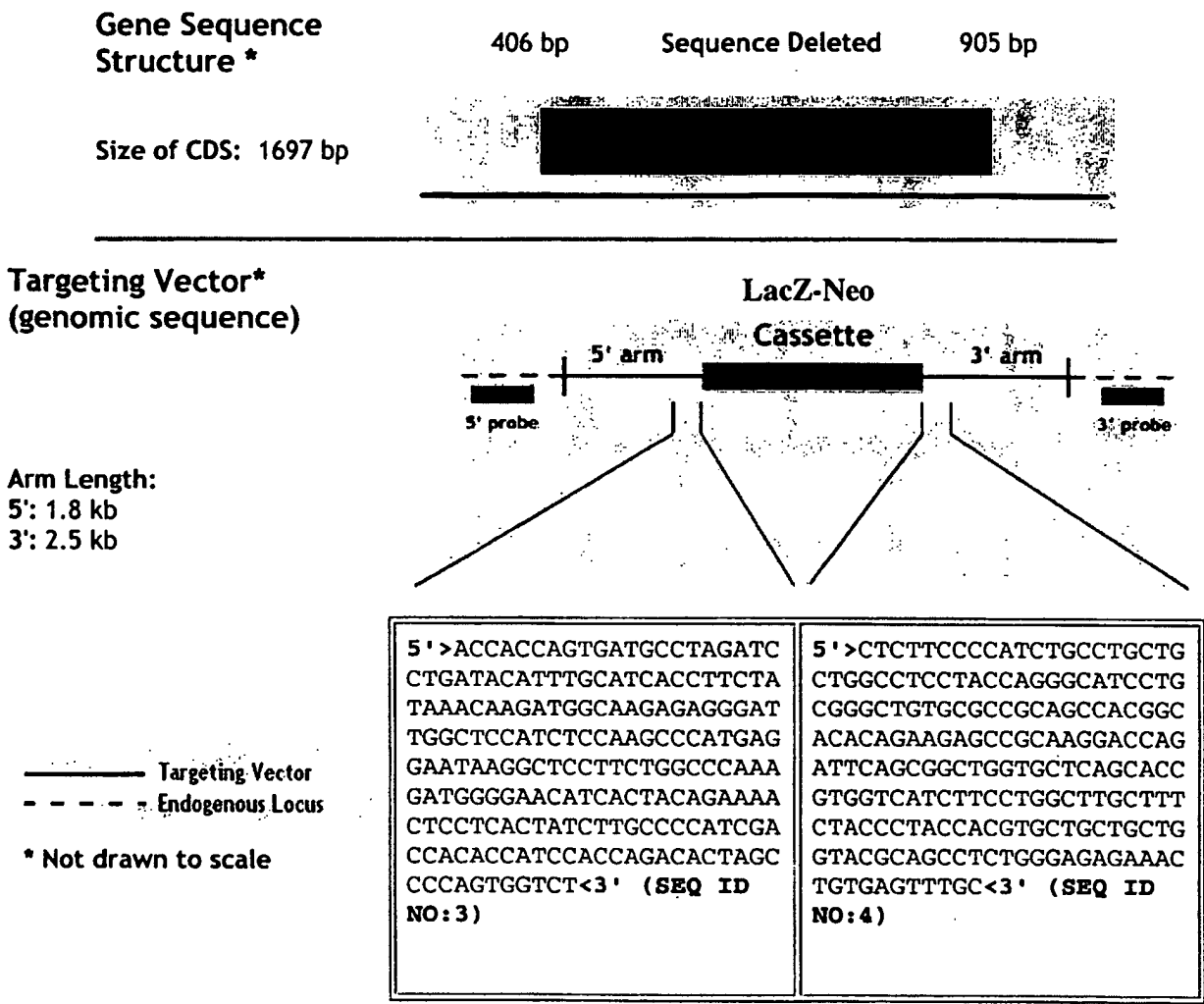


FIGURE 4

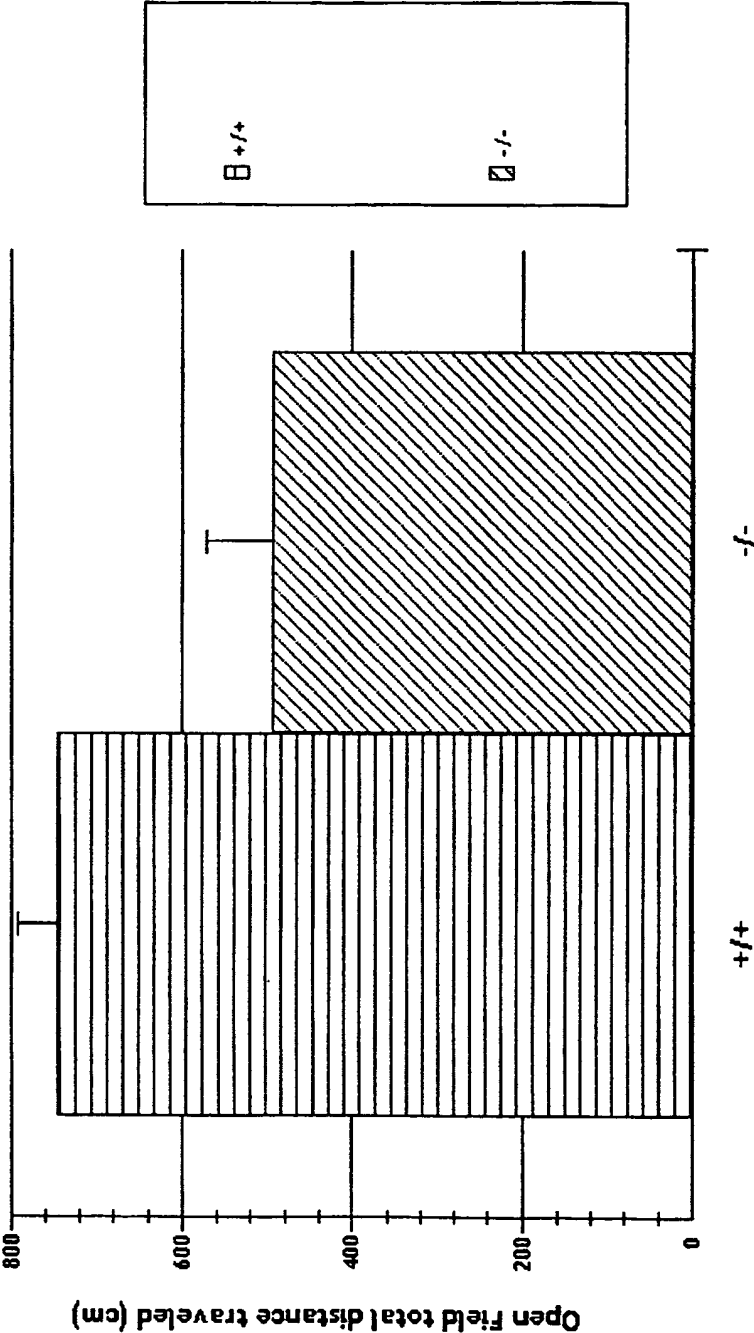


FIGURE 5

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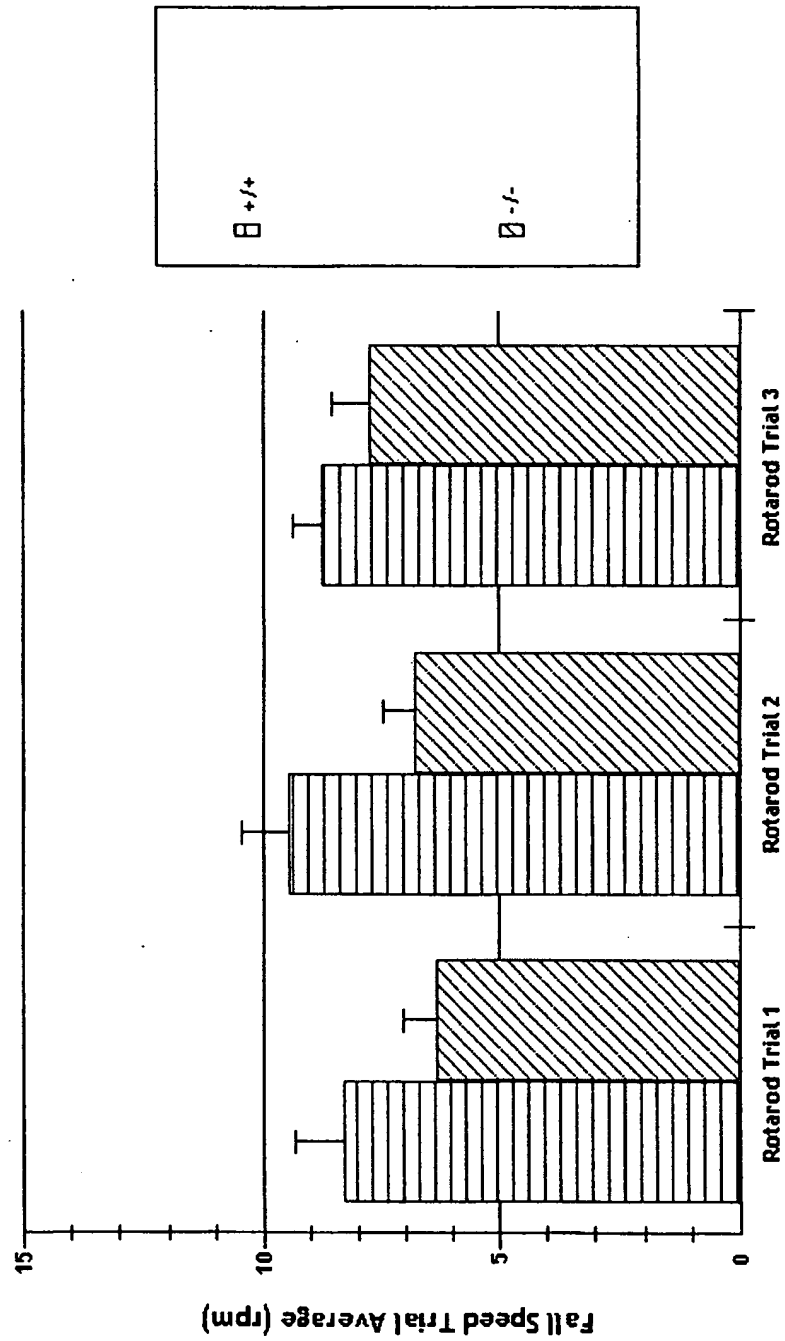


FIGURE 6

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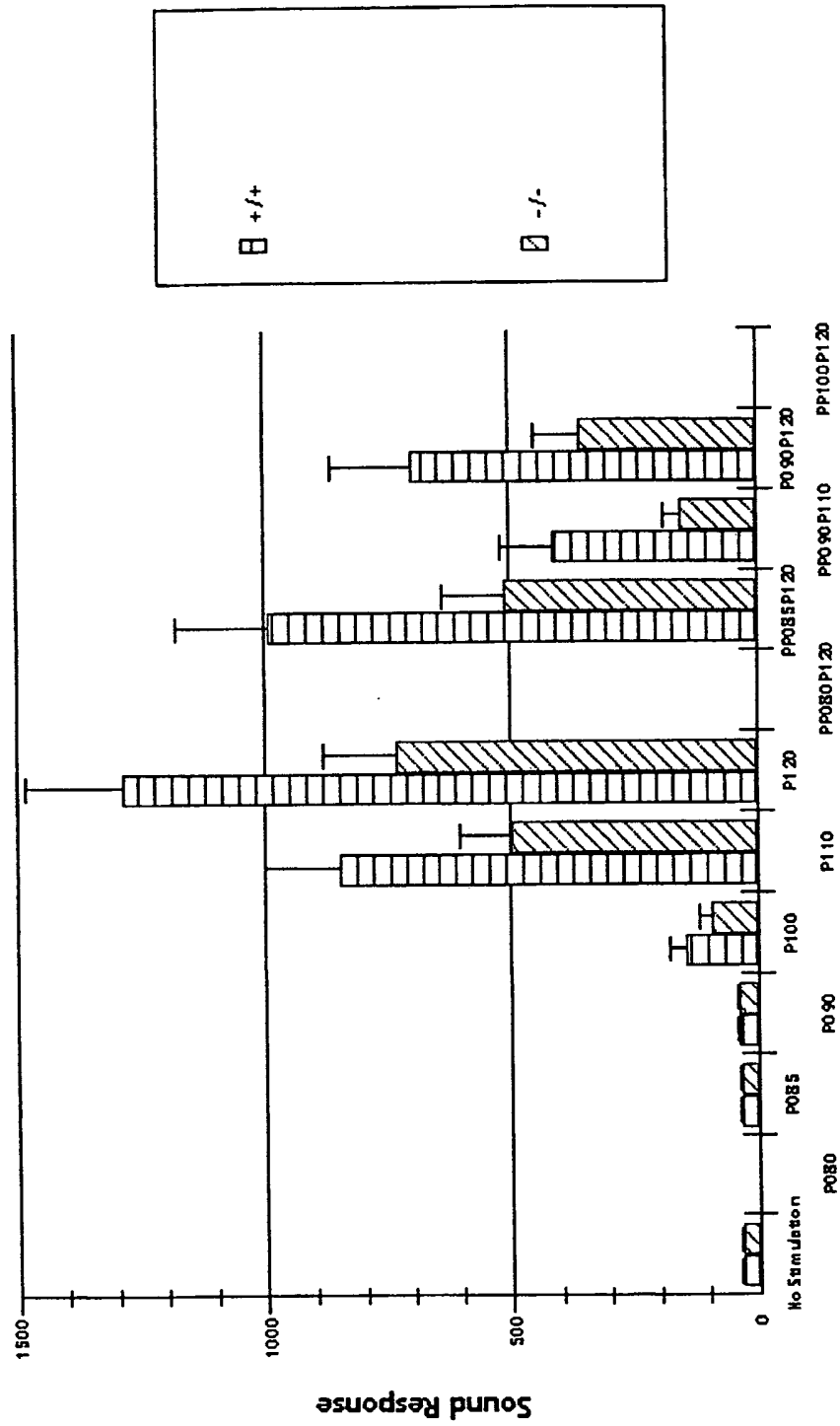


FIGURE 7



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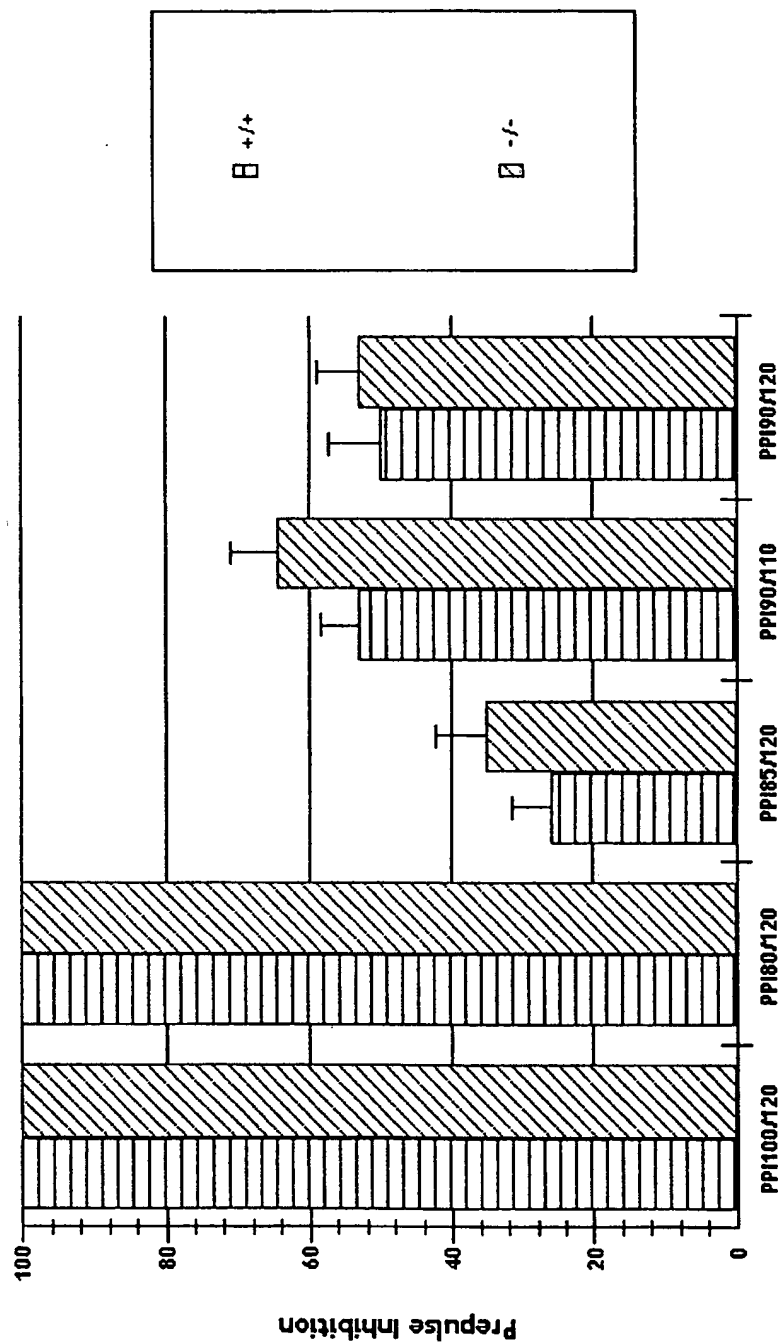


FIGURE 8

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